

# 10 Thyroid Pathology

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## 10.1 Normal Thyroid

The normal thyroid is a bilobed gland, connected by an isthmus. It is encased by a thin capsule that does not strip easily and contains sizable venous channels. The weight of normal thyroid in the United States ranges from 10 to 20 g. The follicle is the functional unit of the thyroid and averages about 20  $\mu\text{m}$  in diameter [1,2,3,4]. A thyroid lobule consists of 20–40 follicles bound together by a thin sheath of connective tissue and supplied by a lobular artery [3,5]. The thyroid follicles are formed by a single layer of low cuboidal epithelium. The nucleus of the follicular cell is round to ovoid in shape; it is usually centrally placed with an inconspicuous nucleolus. The follicle is enveloped by a basal lamina and is surrounded by numerous capillaries and lymphatics [5,6]. The follicular lumen contains colloid, partly composed of thyroglobulin, which is evenly applied to the luminal cell borders. Calcium oxalate crystals are common in the colloid of adults.

Electron microscopy demonstrates that the normal flat to low cuboidal follicular cells interdigitate and overlap one another, and that they are intimately related to the capillaries that surround the follicle; microvilli on the apical surface are numerous near the cellular margins [6,7].

C cells are intrafollicular and are seen next to the follicular cells and within the basal lamina that surrounds each follicle of the normal gland. C cells are most numerous in the central portions of the middle and upper thirds of the thyroid lobes [3]. They are believed to originate from the C cells that arise from the neural crest and migrate with the ultimobranchial body into the thyroid. C cells are typically more numerous in thyroids of infants as compared to adult glands [8,9]. They are polygonal to spindle shaped, have “light” or low density, cytoplasm, and contain numerous membrane-bound cytoplasmic granules containing calcitonin. A small number of C cells (or cells similar to them) contain somatostatin and can increase in number in some patients [10–13].

C cell aggregates can be sizeable (hyperplastic) in some adults without any known endocrinologic

abnormality [14]. C cell hyperplasia is defined as consisting of more than 40 C cells/cm<sup>2</sup> and the presence of at least three low-power microscopic fields containing more than 50 C cells [15]. The small solid cell nests of ovoid to spindle epidermoid cells in thyroid are also considered to be of ultimobranchial origin [15]. Typically, the nests have about the same distribution in the thyroid lobes as the C cells [16,17]. The term “mixed follicles” [18] applies to follicles which are lined by follicular cells and epidermoid cells (and sometimes C cells) and contain both colloid and mucoid material. The ultimobranchial structures probably also give rise to a small proportion of normal thyroid follicles [18].

Oxyphil cells (oncocytes, Askanazy cells, Hürthle cells) are altered/metaplastic follicular cells; they are enlarged, have granular eosinophilic cytoplasm, and have large, hyperchromatic, or bizarre nuclei [19]. The cytoplasm is filled with enlarged mitochondria. They are common in longstanding Graves’ disease, autoimmune thyroiditis, thyroids affected by radiation, follicular-derived neoplasms, and some adenomatous nodules [19–21].

Small aggregates of lymphoid cells in the thyroid stroma can be seen in normal thyroid gland [22]. Also present in the interstitial tissue are antigen-presenting dendritic cells; these are sparse in the normal gland but are increased in autoimmune thyroid disease [23,24].

## 10.2 Developmental Variations

The thyroglossal tract extends in the midline from the foramen cecum at the base of the tongue to the isthmus of the normal gland [25]. The tract consists of connective tissue, the thyroglossal duct, lymphoid tissue, and thyroid follicles; it is attached to and may extend through the center of the hyoid bone and is intimately related to the surrounding skeletal muscle. Thyroid tissue may persist at the base of the tongue and in some patients may be the only thyroid present [25,26]. The thyroglossal duct is typically lined by ciliated pseudostratified epithelium. If the duct is traumatized or infected, the epithelium may undergo alteration to transitional or squamous type, or maybe totally be replaced by fibrous tissue. Foreign body reaction and chronic inflammation may be conspicuous. If fluid accumulates in part of the thyroglossal duct, a thyroglossal cyst may develop [3,27,28].

Any type of diffuse thyroid disease can involve lingual thyroid and the thyroid tissue along the thyroglossal tract [28–30]. In rare instances portions of thyroglossal duct are included within the thyroid

gland proper and, rarely, can serve as the origin of an intrathyroidal cyst [25]. Parathyroid glands, thymic tissue, small collections of cartilage, and glands lined by ciliated cells may be seen in normal thyroids, presumably related to defective development of the branchial pouches [31–33].

Because of the intimate relationship that exists in the embryo between the immature thyroid tissue and the adjacent developing skeletal muscle, strips of striated muscle are occasionally included within the thyroid [34–36].

Thyroid tissue can be found in close proximity or within the perithyroidal skeletal muscle. Such collections of thyroid tissue are particularly prominent when the gland is hyperplastic or is affected by chronic lymphocytic thyroiditis; these should not be confused with carcinoma [34,37].

Groups of thyroid follicles in lateral cervical lymph nodes always represent metastatic carcinoma (papillary carcinoma) [34,37,38]. A few experienced pathologists state normal thyroid follicles rarely occur in cervical lymph nodes [39]. Hence normal thyroid tissue lying only within the capsule of a midline node may represent an embryologic remnant and not metastatic cancer [39,40].

## 10.3 Goiter

Goiter is a diffuse or nodular enlargement of the gland usually resulting from a benign process or a process of unknown origin [41–43]. When there is a deficiency of circulating thyroid hormone because of inborn errors of metabolism, iodine deficiency, or goitrogenic agents, and if the hypothalamic-pituitary axis is intact, production of thyroid-stimulating hormone (TSH; thyrotropin) is increased; consequently, cellular activity and increased glandular activity and glandular mass result in an attempt to achieve the euthyroid state [43–45].

Worldwide, the most common cause for a deficient output of thyroid hormone is an inadequate amount of iodine in the diet, leading to iodine-deficiency goiter (endemic goiter) [46,47]. Other causes of hyperplasia include inborn errors of thyroid metabolism (dyshormonogenetic goiter) [48,49], dietary goitrogens, and goitrogenic drugs and chemicals [50–53].

The pathologic changes of simple non-toxic goiter include one or more of the following: (1) hyperplasia, (2) colloid accumulation, and (3) nodularity [41,54,55]. Hyperplasia represents the response of the thyroid follicular cells to TSH, other growth factors, or to circulating stimulatory antibodies [34,55,56]. The hyperplasia may compensate for thyroid hormonal

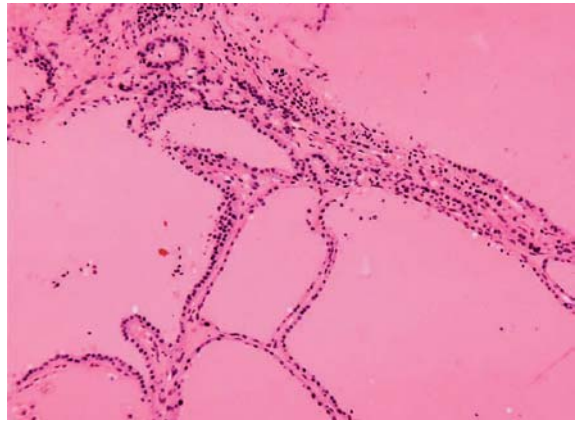
deficiency, but in some cases, even severe hyperplasia does not lead to sufficient hormonal output to avoid development of hypothyroidism.

If the deficiency of thyroid hormone occurs at birth or early in life, cretinism or juvenile myxedema may result, even though the gland is enlarged and hyperplastic; this is especially likely when an inborn error of thyroidal metabolism is present [57,58]. A hyperplastic gland is diffusely enlarged, and not nodular [34,41,56].

Thyroid follicles are collapsed and contain only scanty colloid. The follicular cells are enlarged and columnar in shape with nuclear enlargement, hyperchromasia, and even pleomorphism. When the hyperplastic stage is extreme and prolonged, there may be confusion with carcinoma because of the degree of cellularity and the presence of enlarged cells. Because of follicular collapse and epithelial hyperplasia and hypertrophy, papillary formation can occur [59]. This pattern occurs most often in untreated dyshormonogenic goiter [48]. The recognition of the benign nature of this process is possible because of its diffuse nature [59], unlike carcinoma, in which the tumors grow as localized groups of abnormal cells with a background of non-neoplastic parenchyma.

Thyroid follicles may not remain in a state of continuous hyperplasia but instead undergo a process called involution, with the hyperplastic follicles reaccumulating colloid. The epithelium becomes low cuboidal or flattened and resembles that of the normal gland. The gland is diffusely enlarged, soft, and has a glistening cut surface because of the excess of stored colloid. In addition to large follicles filled with colloid, there are foci in the gland where hyperplasia is still evident (Fig. 10.1). This phase of non-toxic goiter is often termed colloid goiter [60,61].

Patients with long-standing thyroid disorders associated with deficiency of circulating thyroid hormone typically develop nodular goiters that result from overdistention of some involuted follicles, and persistence of the zones of epithelial hyperplasia. The new follicles form nodules and may be heterogeneous in their appearance, in their capacity for growth and function, and in their responsiveness to TSH. The vascular network is altered through the elongation and distortion of vessels leading to hemorrhage, necrosis, inflammation, and fibrosis. These localized degenerative and reparative changes produce some nodules that are poorly circumscribed, and others that are well demarcated and resemble true adenomas (adenomatous goiter) [62,63]. Because the nodules distort vascular supply to some areas of the gland, some zones will contain larger than normal amounts of TSH and/or iodide and others will have relative



**Fig. 10.1** Thyroid follicles lined by low cuboidal epithelium and expanded by thin colloid consistent with colloid goiter

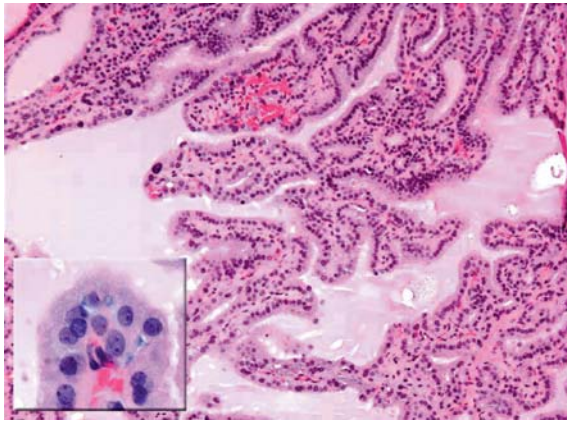
TSH and/or iodide deficiency. Growth of goiters therefore may be related to focally excess stimulation by TSH, stimulation by growth factors, focally abnormal iodide concentration, growth-promoting thyroid antibodies, and poorly understood intrathyroidal factors [47].

Nodular goiter is essentially a process involving the entire gland, but the nodularity may be asymmetric, and individual nodules within the same gland may vary greatly in size. If one nodule is much larger or more prominent than the others (dominant nodule), distinguishing it from a true neoplasm (such as adenoma) may not be possible [37,63]. Several studies have shown that about 70% of dominant nodules in nodular goiter are indeed clonal proliferations [64,65]. The formation of cysts, hemorrhage, fibrosis, and calcification further complicates the assessment of the gland [34,37].

The heterogeneity of the generations of replicating follicular cells, in response to outside stimuli, functional capacity, and rate of growth, forms groups of cells that are hyperfunctional or autonomous, or both. These form “hot” nodules that may cause hyperthyroidism (Plummer’s disease) [66].

### 10.3.1 Graves’ Disease

This disorder is also termed diffuse toxic goiter; it is characterized by diffuse enlargement of the thyroid up to several times normal size. The capsule is smooth and the gland is hyperemic. The cut surfaces are fleshy and lack normal translucence because of loss of colloid. If the patient is untreated, the microscopic appearance shows cellular hypertrophy and hyperplasia [34,67]. The follicular cells are tall columnar and are arranged into papillary formations that extend into



**Fig. 10.2** Graves' disease, papillary hyperplasia. Cells lining the papillae show eosinophilic cytoplasm and round nuclei with even chromatin pattern (*inset*)

the lumina of the follicles (Fig. 10.2). Blood vessels are congested. At the ultrastructural level, microvilli are increased in number and elongated, the Golgi apparatus and endoplasmic reticulum are enlarged, and mitochondria are numerous [56,67]. Lymphoid infiltrates are seen between the follicles, ranging from minimal to extensive. T cells predominate among the epithelial cells (cytotoxic suppressor cells) and in the interstitial tissue (helper inducer cells) where there are no lymphoid follicles [68]. B cells are numerous in the lymphoid follicles. Class II major histocompatibility complex antigens are expressed on the epithelial cells, and these epithelial cells induce the proliferation of T cells, helping to perpetuate the process [68–71].

Lymphoid hyperplasia may occur elsewhere in the body: thymus, lymph nodes, and spleen [72,73].

Because nearly all patients now receive antithyroid medication before surgery, the glands can display varying degrees of involution. In some cases they appear almost normal except for numerous large follicles filled with colloid. A few papillae may remain. The hyperemia is notably decreased, especially if there has been preoperative administration of iodide [63]. If the patient has only been treated for symptoms, i.e., with beta-blockers, the histology of the gland resembles that of the untreated state [74,75].

If hyperplasia continues for many months or several years, oxyphilic/oncocytic metaplasia of the cells begins to occur, the amount of stroma increases in an irregular fashion, and nodularity develops, just as in euthyroid goiter. If the process subsides spontaneously or because of the maintenance on antithyroid medication, the involution may be remarkably complete or irregular (with some foci of hyperplasia evident) [74,75].

In some patients the lymphocytic infiltration is very prominent and resembles the gland affected by chronic lymphocytic thyroiditis [34].

### 10.3.2 Dyshormonogenetic Goiter

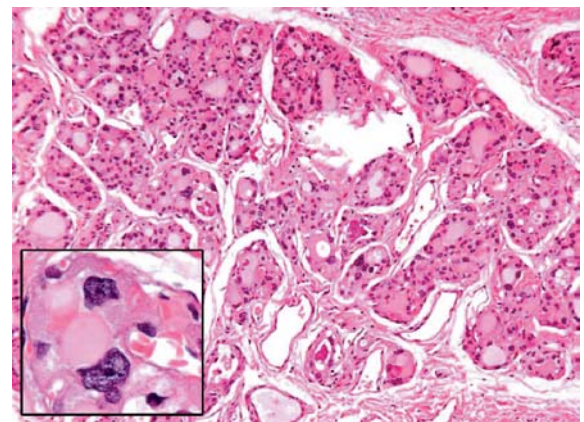
When an inborn error of thyroid metabolism exists, and a sufficient amount of circulating thyroid hormone is not available, the normal physiologic response of the pituitary to increase TSH causes a larger, more active thyroid that may or may not be able to produce enough hormones to reach a euthyroid state. The prolonged and marked TSH stimulation leads to an enlarged and nodular thyroid; microscopically there is enlargement of follicular cells, virtual absence of colloid, and increased stroma [49,76].

Large follicular cells with bizarre, hyperchromatic nuclei may be numerous. The enlarged gland, the bizarre cells, and the cellular nodules have at times been mistaken for carcinoma [48] (Fig. 10.3). Cancer can occur in a dyshormonogenetic goiter, but it is very rare [48,77].

### 10.3.3 Iatrogenic and Related Hyperplasias

Chronic ingestion of excess iodide, for whatever reason, occasionally leads to diffuse hyperplasia. Small nodules with papillary formations may be numerous. Infiltration of lymphocytes may occur [78].

About 3% of patients given lithium salts for a prolonged period develop goiter or hypothyroidism, or both. Patients so treated have been reported to have diffuse hyperplasia with considerable cellular and nuclear pleomorphism [79].



**Fig. 10.3** Dyshormonogenetic goiter. Variably sized aggregates of follicular cells and enlarged pleomorphic nuclei (*inset*)

Bromide ingestion may lead to hypothyroidism because of loss of iodide from the gland. This leads to hyperplastic C cells, foci of papillary proliferation, and loss of colloid [80].

## 10.4 The Thyroiditides

Although occasionally presenting as nodules or asymmetric enlargement of the gland, thyroiditis commonly involves the thyroid diffusely.

### 10.4.1 Acute Thyroiditis

Acute thyroiditis is rare and is almost always due to infection, although acute thyroiditis may be encountered in the thyroid shortly following radiation exposure [81,82]. The disease is most commonly encountered in malnourished children, elderly debilitated adults, immunocompromised individuals, or in otherwise healthy patients following trauma to the neck [81,83]. Most patients present with painful enlargement of the gland. Microscopically acute inflammation with microabscess formation is present. Microorganisms may be seen. A variety of organisms cause thyroiditis including bacteria, fungi, and viruses [84].

### 10.4.2 Granulomatous Thyroiditis

Granulomatous subacute thyroiditis, also referred to as non-suppurative thyroiditis or de Quervain's disease, is a rare entity that usually presents in women and has been associated with HLA Bw35 [85]. The changes seen in the gland are most likely due to the response of the thyroid to systemic viral infection [86–88]; some authors suggest that it represents actual viral infection of the gland. Most patients with subacute thyroiditis recover without any permanent damage to the thyroid. However, some studies have reported end stage hypothyroidism in 5–9% of patients [89]. Microscopically, early in the disease, there is loss of the follicular epithelium and colloid depletion. The inflammatory response, composed initially of polymorphonuclear leukocytes and even microabscesses, progresses until lymphocytes, plasma cells, and histiocytes become the major inflammatory cells. A rim of histiocytes and giant cells replaces the follicular epithelium. A central fibrotic reaction occurs [90]. Recovery is associated with regeneration of follicles from the viable edges of the involved areas [91].

### 10.4.3 Palpation Thyroiditis

Palpation thyroiditis (multifocal granulomatous folliculitis) is found in 85–95% of surgically resected thyroids, and probably represents the thyroid's response to minor trauma. The histologic features of this lesion include multiple isolated follicles or small groups of follicles that show partial or circumferential loss of epithelium and replacement of the lost epithelium by inflammatory cells, predominantly macrophages [92,93].

### 10.4.4 Autoimmune Thyroiditis

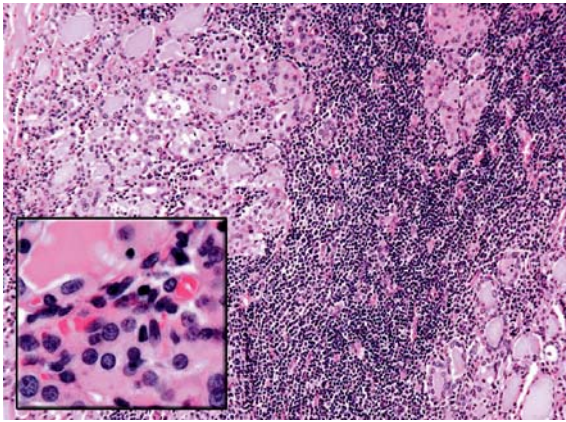
Common synonyms for autoimmune thyroiditis include Hashimoto's thyroiditis, lymphocytic thyroiditis, and struma lymphomatosa [94]. The disorder, most common in women, encompasses a spectrum of clinical and pathologic changes, ranging from an absence of symptoms of thyroid dysfunction to hypothyroidism and rarely, hyperthyroidism, from a large goiter to an atrophic gland, and from scattered clusters of infiltrating lymphocytes to extensive chronic inflammation and scarring with almost complete loss of follicular epithelium [94,95].

Various circulating antithyroid antibodies and other immune phenomena occur, including in situ immune complex deposition and basement membrane changes in the gland and expression of major histocompatibility complex antigens on the thyroid cells [96,97]. The thyroiditis may be found in the same families in which idiopathic hypothyroidism and Graves' disease are common. It may follow typical Graves' disease [98].

The hyperthyroid variant of autoimmune thyroiditis is closely related to Graves' disease and may be almost identical in its gross and microscopic appearance to the latter condition, suggesting that this variant may indeed be Graves' disease [99].

The presence of lymphoid cells in the substance of the thyroid parenchyma probably reflects an abnormal immunologic state. However, the interrelationships among classic chronic thyroiditis, its variants, and "non-specific" thyroiditis are problematic [34]. The morphologic and immunopathologic overlap between non-specific lymphocytic thyroiditis and Hashimoto's disease suggest that they represent a spectrum of autoimmune injury [34,94,100].

In Hashimoto's thyroiditis the gland is firm and symmetrically enlarged weighing from 25 to 250 g [94]. The thyroid has a tan yellow appearance attributed to the abundant lymphoid tissue. The thyroid



**Fig. 10.4** Chronic lymphocytic thyroiditis (Hashimoto's thyroiditis). Oncocytic follicular cells (*inset* showing high power) arranged in nodular pattern with a concomitant lymphocytic infiltrate

follicles are small and atrophic. Colloid appears dense or may be absent. Follicular cells are metaplastic and include oncocytic (Hürthle cell), clear cell, and squamous types. In the stroma and in atrophic follicles, a lymphoplasmacytic infiltration with well-developed germinal centers is found (Fig. 10.4). Variable degrees of interlobular fibrosis are seen [34,94]. The lymphocytic infiltrate is composed of both T and B cells in an almost 1:1 ratio; this differs from the peripheral blood, which shows T cell predominance [101–103]. T lymphocytes within the thyroid are predominantly suppressor type [104,105], whereas the peripheral blood of these patients contains mostly helper T cells. The B cells are usually of the IgG kappa subclass [103].

Patients with Hashimoto's thyroiditis are at increased risk of neoplasia with the most common malignancy being malignant lymphoma, B cell type [106,107]. In addition, patients with Hashimoto's disease may be prone to the development of plasmacytomas within the gland [107]. A peculiar variant of mucoepidermoid carcinoma known as sclerosing mucoepidermoid carcinoma with eosinophilia has been recognized in patients with Hashimoto's disease [108].

#### 10.4.5 Chronic Lymphocytic Thyroiditis Classification

Mizukami et al. established a new classification of chronic lymphocytic thyroiditis [94]. This classification is useful because it allows one to see that the mere presence of lymphocytes in the thyroid does not

indicate autoimmune disease. They basically divided their patients into four groups:

1. *Chronic thyroiditis, oxyphilic*. This group contains patients with classic Hashimoto's disease histology.
2. *Chronic thyroiditis, mixed*. This group shows less of an infiltrate than group 1 with minimal fibrosis. Patients demonstrate either eu-, hyper-, or hypothyroidism.
3. *Chronic thyroiditis, hyperplastic*. This group shows glandular hyperplasia associated with only a small lymphocytic reaction. Most patients are hyperthyroid.
4. *Chronic thyroiditis, focal*. This group shows only a focal lymphocytic reaction and most patients are euthyroid.

#### 10.4.6 Fibrosing Variant of Hashimoto's Thyroiditis

The fibrous or fibrosing variant of Hashimoto's thyroiditis comprises approximately 10–13% of all cases of Hashimoto's disease. Pathologically, the thyroid architecture is destroyed with severe follicular atrophy, dense keloid-like fibrosis, and prominent squamous or epidermoid metaplasia of the follicular epithelium [109,110]. Surgery in this setting can be extremely difficult.

#### 10.4.7 Painless/Silent Thyroiditis

Painless thyroiditis is an autoimmune disease that causes painless enlargement of the thyroid gland along with brief hyperthyroidism followed by hypothyroidism. It can occur in the postpartum period and is termed postpartum thyroiditis [111–113]. Most cases show follicular disruption and lymphocytic infiltration, but stromal fibrous and oxyphilic changes are rare [111].

#### 10.4.8 Focal Non-specific Thyroiditis

Lymphocytic infiltration of the thyroid is found more frequently at autopsy and in surgical specimens since the addition of iodide to the water supplies of the United States about 60 years ago [114,115]. It has been suggested that iodide (iodine) may combine with a protein, act as an antigen, and evoke an immune response localized to the thyroid gland [116]. Postmortem studies indicate an incidence of focal

lymphocytic thyroiditis of about 15–20% in women and rarely in men [114]. These cases show focal aggregates of lymphocytes, occasional germinal center formation, but oncocytes are rarely present. Follicular atrophy is also rare [100].

#### 10.4.9 Riedel's Thyroiditis

Riedel's thyroiditis (Riedel's disease, invasive fibrous thyroiditis, Riedel's struma) has been incorrectly included among the thyroiditides [117]. This is really not a disorder of the thyroid but one that involves the thyroid as well as other structures in the neck or even systemic structures; sclerosing mediastinitis, retroperitoneal fibrosis, pseudotumor of the orbit, and sclerosis of the biliary tract (sclerosing cholangitis) [118–121]. Riedel's disease is an extremely rare entity with an incidence of 0.05% of surgical thyroid diseases and showing a female predominance [122]. Most patients are euthyroid, although hypothyroidism and hyperthyroidism have been reported [117,122].

Descriptions of the thyroid range from stony hard to woody fixed (“ligneous” thyroiditis). Histologically, the involved portions of the gland are destroyed and replaced by keloid-like fibrous tissue associated with lymphocytes and plasma cells [122,123]. The fibrous tissue extends into muscle, nerves, and fat, and entraps blood vessels. In about 25% of cases, the parathyroid glands are also encased [123,124]. There is an associated vasculitis (predominantly a phlebitis) with frequent thrombosis [125].

Quantitative studies of the immunoglobulin-containing cells in fibrous Hashimoto's thyroiditis show that cells containing kappa light chains outnumber lambda-containing cells (64% versus 36%) whereas in Riedel's disease lambda-containing cells comprise >70% of the immunocyte population. In Hashimoto's thyroiditis, IgA cells make up about 15% of the lymphocytes, whereas IgA-containing plasma cells comprise about 45% of the immunocyte population in Riedel's disease. The immunologic evaluation supports the separation of the distinctive Riedel's lesion from other thyroiditides [126].

#### 10.4.10 Combined Riedel's Disease and Hashimoto's Thyroiditis

In rare instances the thyroid gland can show features of both Riedel's disease and Hashimoto's thyroiditis. The histologic picture resembles Riedel's disease, whereas the serology shows thyroglobulin and micro-

somal antibodies seen in Hashimoto's thyroiditis [127].

### 10.5 Amiodarone Injury with Thyrotoxicosis

Administration of amiodarone may cause thyrotoxicosis, primarily due to the large quantity of iodine in the drug [78,128]. Tissue changes are usually focal. Groups of follicles contain degenerated follicular cells (with granular or vacuolated cytoplasm); some follicles have lost follicular cells, and there is partial or complete loss of colloid. Zones of fibrosis are evident. The intervening thyroid tissue is normal [129].

### 10.6 Miscellaneous Disorders

#### 10.6.1 Radiation Effects

Ionizing radiation delivered in small doses to the thyroid glands of infants, children, and adolescents causes a marked increase in the later incidence of benign and malignant neoplasms [130]. Larger doses produce more numerous nodules; many of these nodules are particularly cellular, and some are atypical in their structure and cytologic features, suggesting premalignant characteristics [131]. The cancers that develop after small doses of radiation are mostly papillary carcinomas, are often multicentric or bilateral, and are frequently small [130]. In addition to the nodules and neoplasms that occur, other changes are believed to be more common as well, including focal epithelial hyperplasia (possibly incipient nodules), chronic lymphocytic thyroiditis, oxyphilic metaplasia of follicular cells, and slight fibrosis [132,133].

Large doses of ionizing radiation (e.g., therapeutic radiation for head and neck cancer, or radioiodine therapy) can initially cause vascular injury and follicular cell necrosis. Hemorrhages, edema, and small numbers of the usual inflammatory cells appear. The damaged is healed by scarring and the follicular epithelium can show a mixture of atrophic, hyperplastic, and metaplastic changes [134,135].

#### 10.6.2 Amyloidosis

The thyroid may be involved by primary or secondary amyloidosis. The amyloid deposition may be sufficiently uneven to produce a mass. Such an accumulation must

be differentiated from that occurring in some cases of medullary carcinoma [136,137].

### 10.6.3 Black Thyroid

Prolonged therapy with tetracycline antibiotics, especially minocycline, may cause the accumulation of sufficient pigment in the follicular cells to produce a dark brown to black gland. Much of the pigment is lipofuscin, but part may be a metabolite of the drug. Rarely, there may be interference with thyroid function [138,139].

## 10.7 Neoplasms

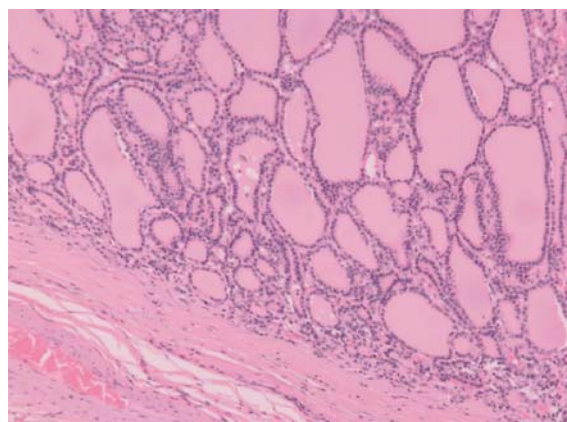
Thyroid neoplasms demonstrate a variety of morphologic patterns that complicate their pathologic interpretation [140]. All neoplasms that arise from thyroid epithelial cells may have some functional capacities. They may respond to TSH and may even produce excessive amounts of thyroid hormones or, if medullary carcinoma, release abnormal quantities of calcitonin or other hormones [34]. Localization of thyroglobulin or calcitonin by immunohistochemistry aids in the classification of unusual thyroidal tumors and in providing definite identification of metastatic thyroid carcinomas [37].

### 10.7.1 Benign Neoplasms

#### 10.7.1.1 Adenomas and Adenomatous Nodules

A follicular adenoma or solitary adenomatous or adenomatoid nodule is defined as a benign encapsulated mass of follicles, usually showing a uniform pattern throughout the confined nodule [35,37]. Follicular adenomas with papillary hyperplasia (some of which are functional) should not be classified as papillary adenomas [141], but as papillary hyperplastic nodules [142]. Adenomas are solitary; indeed, if there are multiple nodules in a lobe or a thyroid gland, it is probably more appropriate to diagnose multinodular goiter with adenomatous change (adenomatous hyperplasia). The features that distinguish histologically between adenoma and adenomatous nodules included encapsulation, uniformity of pattern within the adenoma, and compression of the surrounding gland by the adenoma and its capsule (Fig. 10.5) [143].

Descriptive terms that have been used to delineate the patterns seen in follicular adenomas include macrofollicular, simple, microfollicular, fetal, embryonal,



**Fig. 10.5** Follicular adenoma. Thinly encapsulated follicular patterned lesion and lack of capsular or vascular invasion

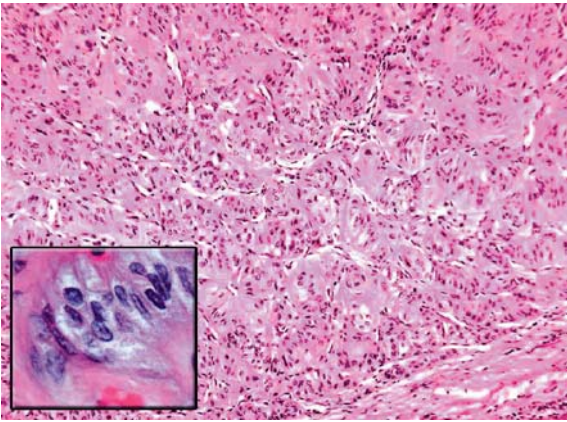
and trabecular [35]. However, since these patterns have no clinical importance, it is not necessary to subdivide thyroid adenomas. Relatively common changes found in adenomas include hemorrhage, edema, and fibrosis, especially in the central portions of the tumor [35]. Calcification may be seen. Lesions that have undergone fine-needle aspiration biopsy may show necrosis, increased mitotic activity, and cellular atypia in the area of the needle tract [144].

Whether or not some solitary follicular nodules have the biologic potential to become carcinoma is unknown; the findings of aneuploid cell populations in 27% of such lesions suggest that some of these may represent carcinoma in situ [145,146]. The solitary follicular lesion that is removed by lobectomy and when adequately studied shows no evidence of invasion, will neither recur nor metastasize [35]. (Enucleation of follicular adenomas or nodulectomy should be condemned as a surgical procedure and considered suboptimal care. The pathologic evaluation of these lesions requires analysis of the tumor capsule–thyroid interface [143].)

### ***Hyalinizing Trabecular Adenoma/Neoplasm of the Thyroid***

The hyalinizing trabecular adenoma is a distinct patterned follicular-derived lesion of the thyroid; i.e., it expresses thyroglobulin, thyroid transcription factor (TTF1) and not calcitonin [147]. Microscopically, these adenomas grow in nests that are surrounded by dense hyaline stroma. The nested histology of the tumor cells is reminiscent of that seen in paragangliomas (thus termed by some authors as paraganglioma-like adenoma of thyroid—PLAT) [148]. The nuclear features of the follicular cells are similar to those seen





**Fig. 10.6** Hyalinizing trabecular neoplasm. Tumor mainly of elongated cells (*inset*) and hyalinized stroma

in papillary carcinoma (Fig. 10.6) [149]. By immunohistochemistry, the cells of hyalinizing trabecular adenoma stain positive for thyroglobulin and cytokeratin 19 and negative for calcitonin, although the presence of other neuroendocrine markers has been described [150].

Some authors have proposed that these adenomas actually represent a variant of papillary carcinoma. This is due to similar nuclear cytology, immunoprofile, and RET oncogene rearrangements in both tumors [151]. However, a benign behavior has so far been described in all cases of hyalinizing trabecular adenoma. Therefore, we believe until metastatic behavior is described in a case of hyalinizing trabecular adenoma with classic histology, these tumors can be designated as hyalinizing trabecular neoplasm as proposed by the World Health Organization [149].

### **Atypical Follicular Adenoma**

The term atypical follicular adenoma includes those follicular tumors that show pathologically disturbing features (spontaneous necrosis, infarction, numerous mitoses, or unusual cellularity), but do not show invasive characteristics on careful examination. The overwhelming majority of the atypical adenomas behave in a benign fashion clinically [152–154].

## **10.7.2 Malignant Neoplasms**

The most common malignant neoplasms of the thyroid origin are the well-differentiated carcinomas of follicular epithelial origin: up to 80% of these are papillary carcinomas [35]. Most non-neoplastic diseases of the thyroid do not seem to be precursors of

malignant diseases, with the exception that autoimmune thyroiditis may predispose to malignant lymphoma [37]. Anaplastic carcinomas have often arisen in goitrous thyroids, and careful examination of the resected tissues has frequently demonstrated benign tumors or well-differentiated carcinomas in close association with the anaplastic neoplasm. Such findings have led to suggestions that the benign tumor or low-grade carcinoma can “transform” into the anaplastic carcinoma [155].

### **10.7.2.1 Papillary Carcinoma**

This is the most common malignant tumor of the gland in countries having iodine-sufficient or iodine-excess diets, and comprises about 80% of thyroid malignancies in the United States [156–158]. Papillary thyroid carcinoma (PTC) clinically behaves in an indolent fashion and carries an excellent prognosis (>90% survival at 20 years) [158]. It invades lymphatics leading to multifocal lesions and to regional lymph node metastases [156,158]. Venous invasion rarely occurs and metastases outside the neck are unusual (5–7% of cases) [159].

Papillary thyroid carcinoma can occur at any age and rarely has been diagnosed as a congenital tumor [160]. Most tumors are diagnosed in patients in the third and fifth decades. Women are affected more than men in ratios of 2:1 to 4:1 [161].

### **Etiologic Factors**

Etiologic factors for PTC are not well established; various cellular and genetic mechanisms/targets have been studied in the development of PTC.

#### *Iodide*

The addition of iodine to the diet in endemic goiter areas in Europe has been associated with a decreased incidence of follicular cancer and an increase in PTC [162,163].

#### *External Radiation*

External radiation probably plays a role in the development of PTC [164,165]. The average time from radiation exposure to tumor development has classically been reported as 20 years; however, development time can be variable [130,165]. The Chernobyl

nuclear accident has induced a great increase in the incidence of PTC in Belarus, Russia, and Ukraine [130]. The increased incidence is seen predominantly in young children. Most reported tumors following this nuclear disaster have been PTC, some of which show aggressive features including extracapsular extension and vascular invasion; however, mortality is extremely low [130,166].

### *Autoimmune Disease*

Many studies indicate that up to one third of PTCs arise in the setting of chronic thyroiditis. Follow-up studies of patients with documented thyroiditis indicate that the tumor that arises much more frequently in these glands is malignant lymphoma, not PTC (see below) [167]. Since papillary cancer and thyroiditis are both common conditions, the possibility of coincidental coexistence is more likely than an etiologic relationship [167,168]. However, loss of heterozygosity for various tumor suppressor genes has been demonstrated in the cytologically atypical areas/nodules in chronic lymphocytic thyroiditis [169]. Thus, a link may exist between chronic lymphocytic thyroiditis and PTC.

### *Hormonal and Reproductive Factors*

Papillary thyroid carcinoma is more common in women than men. Some studies have suggested the role of various hormonal factors in the development of PTC; these include increased parity, late age at the onset of first pregnancy, fertility problems, and oral contraceptives [170]. However, studies of steroid hormone receptors have been disappointing since about 50% of normal thyroid, and benign and malignant nodules can contain estrogen and/or progesterone receptors and smaller numbers of androgen receptors. No correlation with age or gender has been identified [171].

### *Genetic Syndromes*

Papillary carcinomas have been described in patients with familial adenomatous polyposis coli (FAP), Cowden's syndrome, non-polyposis colon cancer syndrome (HNPCC), Peutz Jeghers' syndrome, and ataxia telangiectasia [172–174].

Familial adenomatous polyposis coli is caused by germline mutations of adenomatous polyposis coli

(APC) gene. PTC (>95% of cases) occurs in 12% of patients with FAP; all these patients do show germline mutations of the APC gene, however, somatic mutations or loss of heterozygosity for the APC gene are not found in thyroid tumors. Interestingly, a majority of these tumors do show activation of *ret/ptc1* in thyroid tumors suggesting a possible association between APC and *ret/ptc* in the development of this particular subset of familial papillary carcinoma [172–177].

Cowden's syndrome is characterized by formation of hamartomas in several organs and a high risk of developing breast and thyroid cancer. The genetic locus for Cowden's syndrome has been mapped to chromosome 10q23.3 and is also known as PTEN, which is a protein tyrosine phosphatase and exerts its tumor suppressor effects by antagonizing protein tyrosine kinase activity. Interestingly, PTEN mutation or gene deletion is noted in 26% of benign tumors but only in 6.1% of malignant tumors of the thyroid [178,179].

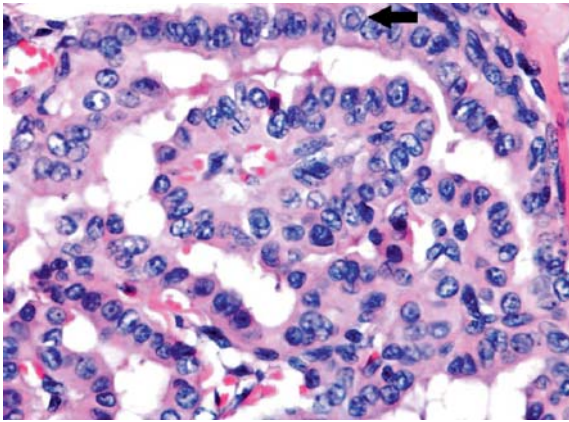
### *Thyroid/Parathyroid Adenomas*

Occasionally, papillary cancers arise in benign nodules or adenomas. It is believed that this is likely to be a random event of location and does not indicate a casual relationship [143]. Several authors have described the association of PTC and parathyroid adenoma and/or hyperplasia [180,181]. Both types of lesions are associated with a history of low-dose external radiation to the neck.

### **Pathology**

The gross appearance of PTC is quite variable. The lesions may appear anywhere within the gland. By definition, clinical papillary carcinomas are >1.0 to 1.5 cm in size often averaging 2–3 cm, although lesions may be quite large. The lesions are firm and usually white in color with an invasive appearance. Tumoral calcification is a common feature. Because of extensive sclerosis, the tumor may grossly resemble a scar. In addition, cyst formation may be observed [34,35,156].

Microscopically, PTC displays papillae containing a central core of fibrovascular tissue lined by one or occasionally several layers of cells with crowded/overlapping oval nuclei. The nuclei of papillary cancer have been described as clear, ground glass, empty, or Orphan Annie eyed. These nuclei are larger and more oval than normal follicular nuclei and contain hypodense chromatin. Intranuclear inclusions of



**Fig. 10.7** Papillary carcinoma, classic type. Tumor cells are arranged in papillary groups and showing chromatin clearing, intranuclear grooves, and inclusions (*arrow*)

cytoplasm are often found. Another characteristic of the papillary cancer nucleus is the nuclear groove (Fig. 10.7) [37,156,157]. Nuclear grooves may be seen in other thyroid lesions including Hashimoto's disease, adenomatous hyperplasia, and diffuse hyperplasia as well as in follicular adenomas (particularly hyalinizing trabecular neoplasm) [182]. Hence, the mere presence of nuclear grooves is not diagnostic for papillary carcinoma.

Psammoma bodies are lamellated round to oval structures that represent the "ghosts" of dead papillae and are formed by focal areas of infarction of the tips of papillae attracting calcium that is deposited upon the dying cells. These are seen within the cores of papillae or in the tumor stroma [35,156]; only rarely are psammoma bodies found in benign conditions in the thyroid [183,184]. Psammoma bodies are found in about 40–50% of cases, but their presence in thyroid tissue indicates that a papillary carcinoma is most likely present somewhere in the gland [35,37]. The finding of psammoma bodies in a cervical lymph node is strong evidence of a papillary cancer in the thyroid [185].

Scattered lymphocytes are common at the invasive edges of the tumor [186,187]. Cyst formation may occur and in fact may be so striking that the diagnosis of PTC is difficult to make particularly if the lesion has metastasized to neck lymph nodes making the distinction clinically from a branchial cleft cyst difficult [34,188,189].

Papillary thyroid carcinoma early in its development invades the glandular lymphatics [34], which accounts for the high incidence of regional node metastases [156]. The tumors can also present multifocally within the same gland [156]. It has been shown

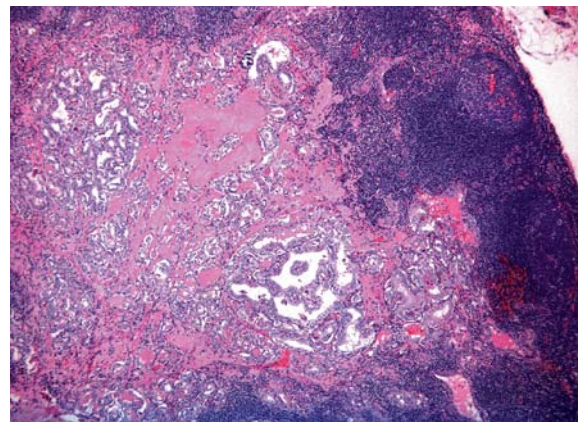
by molecular biology techniques that papillary carcinomas are clonal proliferations [190]. In view of these studies it is believed that multifocality of papillary carcinoma must be due to intrathyroidal lymphatic spread rather than multifocal primary tumors [64,190]. Recent RET/PTC and LOH studies have shown that multifocal papillary microcarcinomas can be separate primaries instead of intraglandular spread from one tumor source [191,192].

Venous invasion can be identified in up to 7% of papillary cancers [193]. Whether this finding alone is predictive of a more aggressive behavior is unclear [194,195].

Regional lymph node metastases are extremely common (50% or more) at initial presentation of usual papillary cancer (Fig. 10.8) [158]. Some patients will present with cervical node enlargement and will have no obvious thyroid tumor [196]. Not infrequently the nodal metastasis will involve one node that may be cystic. The histology of the nodal metastases in papillary cancer may appear papillary, mixed, or follicular. This feature does not adversely affect long-term prognosis [156,157]. Hence, attempts at staging of papillary carcinoma may have minimal clinical significance.

Tumor grading is of no use in this tumor since over 95% of these lesions are grade 1 [194]. In some tumors, either in the primary site or in recurrences, areas of poorly differentiated cancer characterized by solid growth of tumor, mitotic activity, and cytologic atypia can be found. Such lesions have a much more guarded prognosis [197]. Anaplastic change in a papillary cancer can occur, although it is uncommon [155].

Distant metastases of papillary carcinoma to lungs, bones, and brain occur in 5–7% of cases [198]. Despite the presence of multiple metastases, however, survival



**Fig. 10.8** Lymph node with metastatic papillary carcinoma

may still be prolonged, especially if the metastases can be treated with radioiodine [199]. In ordinary papillary carcinoma, death is uncommon [199].

### **Immunohistochemistry**

Immunostaining shows that most papillary cancers express thyroglobulin, TTF1, and not calcitonin [37]. Several reports have been published regarding the use of various immunohistochemical markers that can differentiate papillary carcinoma from other follicular-derived lesions of the thyroid. From an extensive list of these markers the ones that have shown some promise include cytokeratin 19, HBME1, and galectin 3 [200–205]. However, none of these have proven to be specific since all can be expressed in some benign lesions of the thyroid. Therefore, some authors have proposed that diagnosis of PTC by immunohistochemistry should be carried out by using the markers mentioned above in an immunopanel [202,206].

The other markers that have been explored in the diagnosis of PTC include: S100 protein, blood group antigens, estrogen receptors, CD10, CD15, and CD57. The proliferation of markers indicates that no one of them is useful for the daily practice of pathology [207–210].

### **Flow Cytometry**

Although the great majority of papillary thyroid cancers are diploid, the literature suggests up to 20% may show aneuploid or at least non-diploid subpopulations. It has been shown that aneuploid tumors are often associated with a more aggressive clinical course; however, multivariate analysis has not shown that ploidy is an independent prognostic factor [146,157,211].

### **Molecular Pathology**

In the decade since 1995 the literature on thyroid has been focused mainly on the role of various biologic events and genetic determinants in the pathogenesis of various thyroid tumors. Rearrangements of RET gene, known as RET/PTC have been identified in papillary carcinoma of the thyroid [212,213]. The RET protooncogene is normally expressed in cells of neural crest origin and plays a role in kidney and gastrointestinal neuronal development. It is located on chromosome 10q11.2 and cell membrane receptor tyrosine kinase [212,214]. In normal thyroid wild-type

RET is only expressed in C cells and not follicular cells. RET/PTC seen in papillary carcinomas occurs due to fusion of the tyrosine kinase domain of RET to the 5' portion of the various genes. To date more than ten novel types of rearrangements have been described in papillary carcinoma. RET/PTC1 and 3 are the most common forms that occur in sporadic papillary carcinoma. RET/PTC1 is formed by fusion of RET to *H4* and RET/PTC3 occurs due to fusion of RET to *ELE1* gene [214–216].

RET/PTC expression in thyroid follicular cells of transgenic mice leads to development of thyroid tumors with histologic features of papillary thyroid carcinoma [217]. Similarly, transfection of follicular cells in tissue culture by RET/PTC causes the cells to demonstrate nuclear features of papillary carcinoma [218]. The prevalence of RET/PTC in papillary carcinoma varies significantly among various geographic regions; in the United States it ranges from 11% to 43% [216]. In sporadic tumors RET/PTC1 is the most common form of rearrangement (60–70%) followed by RET/PTC3 (20–30%) [216,219]. The other rare forms of RET/PTC rearrangements have been mainly found in radiation-induced papillary carcinomas. Several studies have shown a strong association between radiation-induced papillary carcinoma and expression of RET/PTC; in papillary carcinoma in children affected by the Chernobyl nuclear accident, RET/PTC3 was found to be the commonest form of rearrangement followed by RET/PTC1 [220,221].

Recently it has been shown that RET/PTC expression can also occur in some benign lesions. These include hyalinizing trabecular neoplasm [151], Hashimoto's thyroiditis [222,223], and hyperplastic nodules and follicular adenoma [224].

Several authors have suggested an association between Hashimoto's thyroiditis and PTC; however, others have suggested that this association is most likely incidental since both are common. Recently two independent studies have shown high prevalence of RET/PTC in histologically benign thyroid tissue affected by Hashimoto's thyroiditis; these studies concluded that thyroiditic glands harbor multiple foci of papillary carcinoma that are not identified by histologic examination only [222,223]. However, a more recent study failed to reproduce these results [168].

RET/PTC has been identified in benign thyroid nodules, especially the ones that are seen in patients with a history of external radiation [224]. However, this still remains controversial and needs to be further elucidated by examination of a large cohort of cases.

Activation of the ras oncogene-signaling pathway is considered to be an important mechanism by which human cancer develops. Ras has been shown to regu-

late several pathways that contribute to cellular transformation including the Raf/MEK/ERK pathways. Numerous studies confirm that the Raf/MEK/ERK pathway is a significant contributor to the malignant phenotype associated with deregulated Ras signaling [225,226].

Recently, an activating mutation in the serine/threonine kinase BRAF was described of human PTCs. BRAF-activating mutations in thyroid cancer are almost exclusively the BRAF V600E mutation, and have been found in 29–69% of papillary thyroid cancers, 13% of poorly differentiated cancers, and 10% of anaplastic cancers [226–229]. These data identify that BRAF is an oncogene in human cancer. The high frequency of BRAF mutations in thyroid cancer suggests that inhibition of BRAF activity may represent an important new strategy in the treatment of patients with thyroid cancer.

### Prognostic Factors

Poor prognostic factors in papillary carcinoma include older age at diagnosis, male sex, large tumor size, and extrathyroidal growth [158,199]. Pathologic variables associated with a more guarded prognosis include less differentiated or solid areas, vascular invasion, and aneuploid cell population [194]. Some authors have found varying prognostic factors in males and females. In men, age and presence of gross lymph node metastases were important, while in females age, presence of gross lymph node metastases, tumor size, and the number of structures adhered to the gland were important [199,230].

Some studies have shown that RET/PTC expression in papillary carcinoma can be associated with aggressive biologic behavior; conversely, others have reported that its expression is more commonly seen in slow growing and clinically indolent tumors [231,232]. It is also suggested that different rearrangements of RET/PTC are associated with different biologic behavior. Nikiforov et al. found a significant difference in local recurrence and distant metastases between tumor with RET/PTC1 and RET/PTC3 expression [216]. Cetta et al. reported similar findings [233]. Besides RET/PTC, several other biologic markers have been suggested as prognostic predictors in papillary carcinoma; these include p53, Ki67, cell cycle proteins, proliferating cell nuclear antigen (PCNA), bcl2, cathepsin D, and topoisomerase II [234–238].

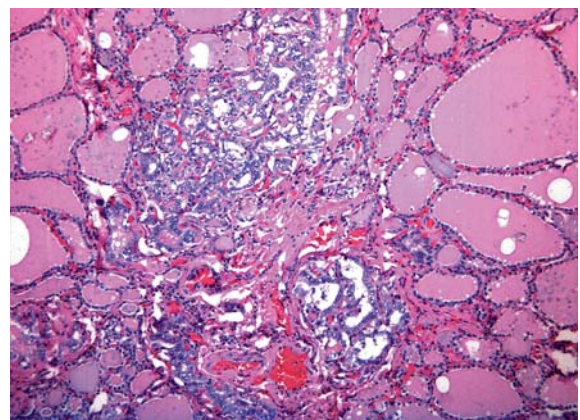
### Subtypes of Papillary Carcinoma

#### *Papillary Microcarcinoma (Occult Papillary Carcinoma)*

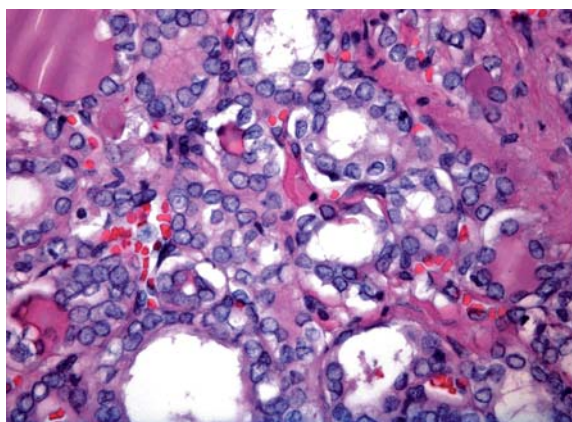
According to the WHO, papillary microcarcinoma is defined as tumor measuring 1 cm or less; however, some experts have also defined tumors measuring up to 1.5 cm as microcarcinomas (Fig. 10.9) [239,240]. These lesions are quite common as incidental findings at autopsy or in thyroidectomy for benign disease or in completion thyroidectomies in patients with a history of carcinoma involving the opposite thyroid lobe [241]. The incidence of these lesions has varied significantly with the study, but papillary microcarcinoma has been reported in up to 36% of carefully sectioned thyroid specimens [241]. Lymph node metastases from papillary microcarcinoma can occur; metastases from lesions less than 0.5 cm have been reported [240,241]. Distant metastases, although very rare, are also documented [242]. Histologically, the tumors may be totally follicular or show papillary areas as well. Sclerosis may be prominent; the lesions infiltrate the surrounding thyroid [34]. A familial form of papillary microcarcinoma has been recognized; these tumors are characterized by multifocality with increase tendency toward vascular and lymphatic invasion, distant metastasis, and even death [243,244]. It is important to recognize that the incidentally found microcarcinoma confined within the thyroid is probably of no clinical importance and should not be overtreated.

#### *Follicular Variant of Papillary Cancer*

The follicular variant of papillary carcinoma is a distinctive papillary carcinoma variant that shows follic-



**Fig. 10.9** Papillary microcarcinoma

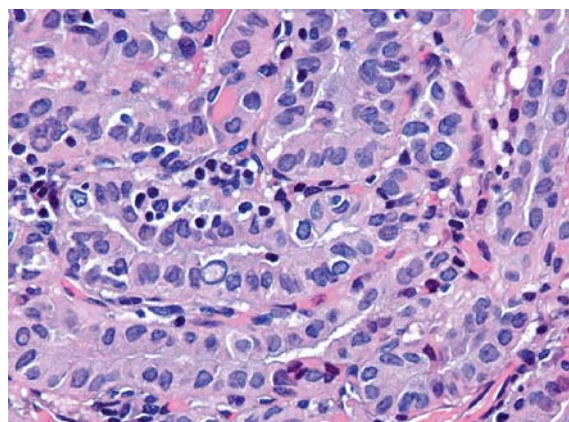


**Fig. 10.10** Follicular variant of papillary carcinoma. Variably sized follicles lined by cells showing nuclear features of papillary carcinoma

ular growth pattern and diagnostic nuclear features of papillary carcinoma (Fig. 10.10) [245,246]. The incidence of this variant is difficult to determine since in the past some of these lesions have been classified as follicular carcinomas or adenomas [247]. Grossly and histologically, the tumor may appear encapsulated [248]. The prognosis of the follicular variant is apparently similar to usual papillary cancer although there may be a greater risk for this variant to metastasize outside the neck and for vascular invasion; regional nodal metastases are less common than in classic papillary cancer [249,250].

Two distinct types of follicular variant are the diffuse and the encapsulated follicular variants. In the diffuse follicular variant, the gland is diffusely replaced by tumor [251]. Lymph node and distant metastases are common in these patients. The prognosis appears to be poor in these patients, although only a handful of cases have been described [252,253].

The encapsulated follicular variant refers to the follicular variant that is characterized by the presence of a capsule around the lesion. These lesions are associated with an excellent prognosis [251]. In some cases the diagnosis of this particular variant of papillary carcinoma can be difficult due to the presence of multifocal rather than diffuse distribution of nuclear features of papillary thyroid carcinoma. Because of this peculiar morphologic presentation, these tumors can be misdiagnosed as adenomatoid nodule or follicular adenoma [143,251]. Some authors have suggested that these tumors be classified as “tumors of undetermined malignant potential” due to their excellent prognosis [254]; however, others have shown that some cases belonging in this category can lead to distant metastasis [249].



**Fig. 10.11** Tall cell variant of papillary carcinoma. Enlarged tumor cells with oncocyctic cytoplasm and nuclear features of papillary carcinoma

#### *Tall Cell Variant*

The tall cell variant is an aggressive variant of papillary carcinoma that tends to occur in elderly patients. These tumors are usually large (>6 cm), extend extrathyroidally, and show mitotic activity and vascular invasion more often than usual papillary cancer. The tall cell variant of PTC consists of tumor cells twice as tall as they are wide and shows eosinophilic cytoplasm; because of this these tumors are referred to as the “pink cell” variant of papillary carcinoma (Fig. 10.11) [255,256]. Dedifferentiation to squamous cell carcinoma has been described in the tall cell variant of PTC [257]. The prognosis for this variant is less favorable than for usual papillary cancer, although it is believed that the poor outcome is secondary to the fact that these tumors are often associated with poor prognostic variables such as older age, extrathyroidal spread, necrosis, high mitotic rate, and distant metastases [258–260].

#### *Columnar Cell Variant*

The columnar cell variant is a rare form of papillary carcinoma [261]. (Some authors believe it is so unusual a tumor that it deserves its own category and should not be placed in the papillary group. The tumor needs to be distinguished from other papillary carcinomas since this lesion is associated with an extremely poor outcome with most deaths occurring within 5 years of diagnosis. Extrathyroidal extension is common as are distant metastases [261–263]. Encapsulated variants, which may have a better prognosis, have been described [264].

### *Warthin-like Variant*

By light microscopy these tumors resemble “Warthin’s tumor” of the salivary gland. These tumors usually arise in a background of lymphocytic thyroiditis and show papillary architecture. Limited follow-up has shown that these tumors in their pure form follow a clinical course similar to conventional papillary carcinoma [265,266].

### *Diffuse Sclerosis Variant*

The diffuse sclerosis variant of papillary carcinoma is rare; it most often affects children and young adults, and may present as bilateral goiter. The tumor permeates the gland outlining the intraglandular lymphatics. The lesions tend to recur in the neck and have a somewhat more serious prognosis than usual childhood papillary cancer. These lesions appear to represent 10% of the papillary carcinomas seen in children exposed to the radioactive iodine released following the Chernobyl accident. While the tumors often show extracapsular extension, distant and nodal metastases, and a decreased disease-free survival when compared to the usual type of papillary carcinoma, mortality is low [267–270].

### *Solid Variant of Papillary Carcinoma*

The solid variant of PTC is most commonly seen in children and has been reported in greater than 30% of patients with papillary carcinoma following the Chernobyl nuclear accident [271,272]. It is important to recognize these lesions as papillary carcinomas and not to classify them as more aggressive tumors such as insular carcinoma (discussed below). The prognosis is controversial with some studies showing outcomes similar to typical papillary carcinoma and other studies showing more aggressive behavior [271,273].

### *Other Variants of Papillary Carcinoma*

Rare variants of papillary cancer in which prognostic data are not well established include the spindle cell variant [274], the clear cell type [275], the oxyphilic (Hürthle cell) variant [275,276], papillary carcinoma with lipomatous stroma [277,278], papillary carcinoma with fasciitis-like stroma [279], and the cribriform variant [280–281]. The last of these is often seen in patients with familial adenomatous polyposis

although it may occur as a sporadic tumor. It is overwhelmingly common in women [175,282].

### **10.7.2.2 Follicular Carcinoma**

Follicular carcinoma comprises about 5% of thyroid cancers; however, in iodide-deficient areas, this tumor is more prevalent making up 25–40% of thyroid cancers [283,284]. The true incidence of follicular carcinoma is difficult to determine since the follicular variant of papillary carcinoma may still be placed into this category [247]. Risk factors include iodine deficiency, older age, female gender, and radiation exposure (although the relationship of radiation to follicular carcinoma is far less strong than with papillary cancer) [162,285]. Clinically, follicular carcinoma usually presents as a solitary mass in the thyroid [283].

Follicular carcinoma has a marked propensity for vascular invasion and avoids lymphatics; hence, true embolic lymph node metastases are exceedingly rare. Follicular carcinoma disseminates hematogenously and metastasizes to bone, lungs, brain, and liver [283,286,287].

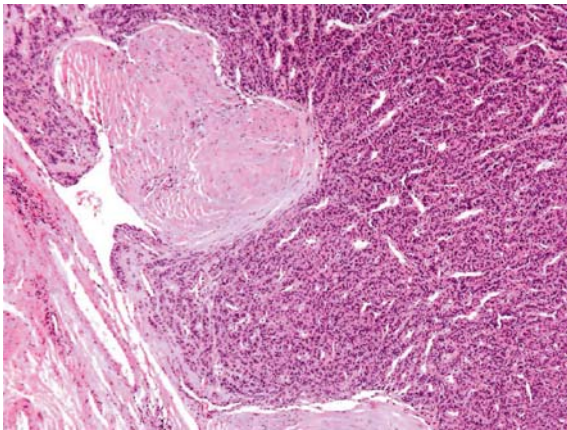
Patients who have follicular carcinoma that is widely invasive fare poorly [284,288]; however, those individual with encapsulated follicular tumors confined to the thyroid enjoy a prolonged survival (greater than 80% at 10 years) [289–292]. Studies using multivariate analysis have identified age >45, extrathyroidal extension, distant metastases, and tumor size >4 cm as independent prognostic factors in follicular carcinoma [286,291,293]. An extremely significant complication that may occur in patients with follicular cancer is transformation into anaplastic cancer; this may occur *de novo* in an untreated follicular lesion, or in metastatic foci [294].

The *widely invasive follicular carcinoma* is a tumor that is clinically and surgically recognized as a cancer; the role of the pathologist in its diagnosis is to confirm that it is of thyroid origin and is a follicular neoplasm. Up to 80% of the patients with widely invasive cancers can develop metastases, and a 50% fatality rate for widely invasive tumors compared with only 3% for those with minimal invasion has been reported.

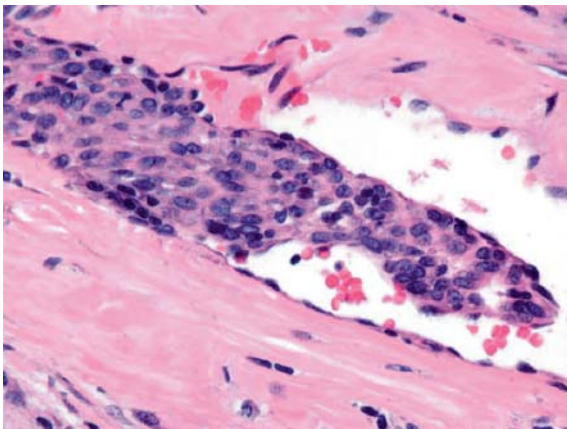
The pathologist can only diagnose the minimally invasive follicular carcinoma by examining well-fixed histologic sections. These lesions are not diagnosable by fine-needle aspiration cytology since the diagnosis requires the demonstration of invasion at the edges of the lesion; therefore, sampling of the center, as in obtaining a cytologic sample, cannot be diagnostic [283–287,295–297].

The *minimally invasive follicular carcinoma* is an encapsulated tumor that grossly resembles a follicular adenoma and only on microscopic examination shows evidence of capsular and/or vascular invasion (Figs. 10.12, 10.13). On microscopic examination, follicular carcinomas most often have a microfollicular pattern and resemble a cellular follicular adenoma. Trabecular or solid patterns are fairly common and often accompany the microfollicular pattern [143,291].

What are the minimum criteria for making this diagnosis? Invasion of the capsule, invasion through the capsule, and invasion into veins in or beyond the capsule represent the diagnostic criteria for carcinoma in a follicular thyroid neoplasm [143,247,291]. The criterion for vascular invasion applies solely and strictly to veins in or beyond the capsule, whereas, the defi-



**Fig. 10.12** Follicular carcinoma. Thickly encapsulated follicular and solid patterned lesion invading into the capsule in a mushroom-shaped growth



**Fig. 10.13** Follicular carcinoma. Tumor embolus within a capsular vessel

inition of capsular invasion is controversial [143,254]. Some authors require penetration of the capsule to diagnose a follicular tumor as carcinoma, while others need tumor invasion through the capsule into the surrounding normal thyroid [254,292]. Is capsular invasion insufficient for the diagnosis of follicular cancer? Distant metastases have been reported in follicular carcinoma diagnosed only on the basis of capsular and not vascular invasion, however, in some cases, metastases were already present at initial diagnosis [290,298]. The presence of vascular invasion is also indicative of malignancy in a follicular tumor. Invasion of vessels within or beyond the lesional capsule is necessary for a definitive diagnosis of vascular invasion [35]. The lesions with vascular invasion should be separated from the minimally invasive follicular carcinomas that show capsular invasion only, because angioinvasive lesions have a greater probability of recurrence and metastasis [251].

In our practice, we use the terms *minimally invasive* and *angioinvasive carcinoma*. The former is applied to those cases that show only capsular or transcapsular invasion, while the latter is used for tumors in which vascular invasion is found with or without capsular invasion. As mentioned above, we propose this distinction based on the belief that angioinvasive tumors have a greater propensity toward distant metastasis.

Similar problems exist in evaluating such lesions by frozen section [299,300]. Some authors recommend that intraoperative assessment of such lesions involves the examination of frozen sections from three or four separate areas of the nodule [301]. This wastes resources and rarely gives useful diagnostic information. The surgeon should have removed the lobe involved by the nodule and if it is a follicular carcinoma that is only minimally invasive, the appropriate therapy has probably already been accomplished. Since a only small number of these lesions will show evidence of invasion at the time of permanent section, i.e., the majority of them are benign, and since overdiagnosis is more dangerous for the patient than is the delay in making a definitive diagnosis [299], we discourage frozen section evaluation of these nodules.

None of the ancillary techniques assist in defining benign from malignant follicular tumors. Ultrastructural, morphometric, and flow cytometric analyses have not helped in distinguishing these lesions [145,302]. About 60% of follicular carcinomas will show aneuploid cell populations [145]. Backdahl analyzed 65 follicular thyroid tumors (26 benign and 39 carcinomas). He noted that of the 20 patients with cancer who survived, 19 had diploid tumors, whereas



17 of 19 patients who died of carcinoma had tumors with aneuploid DNA patterns [303].

All follicular carcinomas express thyroglobulin and show a similar cytokeratin profile to normal thyroid parenchyma. Some authors have shown that HBME1 is exclusively expressed in 90–100% of follicular carcinomas and not adenomas. However, others have reported HBME1 expression in adenomatoid nodules and follicular adenomas [204,210,304,305].

### **Molecular Biology of Follicular Carcinoma**

A specific translocation t(2;3) leads to the expression of PAX8 peroxisome proliferator activated receptor gamma (PPAR gamma) chimeric protein; initial studies by Kroll et al. demonstrated that this translocation is specific to follicular carcinoma [306]. However, follow-up studies employing immunohistochemistry and molecular biology have shown that PPAR gamma expression can occur in some cases of follicular adenoma, follicular variant of papillary thyroid carcinoma, and even benign thyroid parenchyma [307,308]. *Ras* mutations are more frequent in follicular carcinoma as compared to follicular adenoma; some authors have found an association between *ras* mutations and clinically aggressive follicular carcinomas [309–311]. Loss of heterozygosity on chromosomes 10q and 3p can be seen in follicular carcinoma suggesting a role of tumor suppressor genes in its pathogenesis [312,313].

### **Well-differentiated Follicular “Tumors of Undetermined Malignant Potential”**

This designation has been recently proposed in thyroid pathology for follicular patterned encapsulated tumors that have been controversial and difficult to diagnose due to: (1) questionable or minimal nuclear features of papillary thyroid carcinoma or (2) questionable or one focus of capsular invasion that is confined to tumor capsule and does not traverse the entire thickness of capsule and lacks any nuclear features of papillary thyroid carcinoma [254].

This terminology may be extremely helpful to pathologists in the diagnoses of certain follicular patterned lesions; however, these terms are proposed on the basis of data that lack complete clinical follow-up. Therefore, clinicians may find it problematic to establish treatment strategies [143].

### **Oncocytic (Hürthle Cell) Tumors**

Hürthle cells are derived from follicular epithelium and are characterized morphologically by large size, distinct cell borders, voluminous granular cytoplasm, large nucleus, and prominent nucleolus. Ultrastructural studies have shown that the cytoplasmic granularity is produced by huge mitochondria filling the cell [314,315]. Hürthle cells can be found in a number of conditions in the thyroid [nodular goiter, non-specific chronic thyroiditis, longstanding hyperthyroidism, and chronic lymphocytic thyroiditis (Hashimoto’s disease)] [19].

Perhaps no thyroid neoplasm has elicited more confusion or debate than Hürthle cell (oncocytic) neoplasms. Clinicians and pathologists alike have considered that such tumors do not “follow the rules” for histopathologic diagnosis of malignancy. Some authors cite 80% or more of these lesions as benign, whereas others consider all such lesions malignant [316,317]. Over the decade since 1995, studies from numerous institutions throughout the world have shown that oncocytic or Hürthle cell tumors can be divided into benign and malignant categories by careful adherence to strict pathologic criteria [318,319].

Since most Hürthle cell neoplasms are follicular in pattern, the criterion for distinguishing benign from malignant is the same as for follicular neoplasms, i.e., the identification of capsular and/or vascular invasion [318,319]. However, the pathologic criterion for malignancy is met more frequently for tumors composed of Hürthle cells than for their non-Hürthle counterparts. Thus, whereas 2–3% of solitary encapsulated follicular tumors of the thyroid show invasive characteristics, 30–40% of such lesions showing Hürthle cell cytology will show such features [315,318,320]. In addition, whereas true follicular carcinomas of the thyroid rarely, if ever, metastasize embolically to lymph nodes, about 30% of Hürthle cell carcinomas do [34,321].

Most Hürthle cell neoplasms of the thyroid are solitary mass lesions that show complete or partial encapsulation. They are distinguished from the surrounding thyroid by their distinctive brown to mahogany color [34,35,318]. Rarely, a Hürthle cell neoplasm may undergo spontaneous infarction. Extensive infarction may also be seen following fine-needle aspiration biopsy.

The claim that all Hürthle cell neoplasms should be considered malignant or potentially malignant, especially if 2 cm or greater in size, is no longer considered valid. Many studies from the United States and Europe indicate that benign Hürthle cell neoplasms

exist. Size, nuclear atypia, multinucleation, cellular pleomorphism, mitoses, or histologic pattern of the lesion are not predictive of behavior [315,318,319].

By immunohistochemistry, Hürthle cell lesions are positive for thyroglobulin. Carcinoembryonic antigen (CEA) expression has been described in some, but not all series. Hürthle cell lesions are positive for S100 protein [318,322].

DNA ploidy studies have shown aneuploid DNA patterns in biologically and histologically benign Hürthle tumors of the thyroid. These findings do not indicate malignant behavior; however, about 20–50% of Hürthle cell tumors that are histologically malignant and aneuploid are more aggressive biologically and clinically than diploid Hürthle cell cancers [323].

### *Molecular Biology of Hürthle Cell Tumors*

Hürthle cell tumors are biologically different from other follicular-derived tumors. *H-ras* mutations are more frequent in Hürthle cell carcinoma than follicular carcinoma [324,325], and a high percentage of allelic alterations occur as compared to other follicular-derived tumors. A study by Maximo et al. showed that Hürthle cell tumors display a relatively higher percentage of common deletions of mitochondrial DNA as compared to other follicular-derived tumors. In addition, Hürthle cell tumors also showed germline polymorphisms of ATPase 6 gene, which is required for the maintenance of mitochondrial DNA [326].

### **Clear Cell Tumors**

Clear cell change of the cytoplasm can occur in many follicular-derived lesions of the thyroid, thyroiditis, nodules, and neoplasms [278,327,328]. Of greatest importance is the differentiation of clear cell change in follicular thyroid lesions from clear cell renal cell carcinomas metastatic to the thyroid [329]. Immunostains for thyroglobulin are usually helpful in sorting out this diagnostic problem.

### **Poorly Differentiated Carcinoma/ Insular Carcinoma**

This heterogeneous group of malignant thyroid tumors includes carcinomas that are recognizable as originating from follicular epithelium (often with evidence of coexistent papillary or follicular carcinoma),

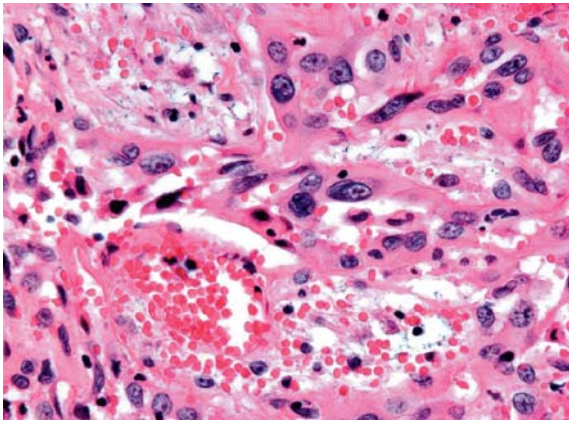
but that have moderate to high rates of mitotic activity, are composed of solid masses or trabeculae of relatively uniform epithelial cells, have tiny follicles present in varying numbers, may contain regions of acute necrosis, and are more aggressive than the usual well-differentiated carcinomas [197]. Included among these lesions are insular carcinoma, columnar cell, tall cell, and trabecular types of papillary cancer, and “poorly differentiated” carcinoma of Sakamoto [330].

Insular carcinoma or poorly differentiated thyroid carcinoma is a follicular-derived carcinoma with a prognosis between well-differentiated thyroid carcinomas (papillary or follicular) and anaplastic thyroid carcinoma. The term “insular” is used to describe the lesion’s histologic growth pattern, which is somewhat “carcinoid-like.” The incidence of this tumor appears to vary with differing geographic locations with incidence as high as 5% described in Italy, while the incidence in the United States is much lower [331].

The lesions are often large, gray-white in color, infiltrative, and show extensive necrosis. Microscopically the tumor is composed of small nests of cells that have a neuroendocrine growth pattern. Necrosis, vascular invasion, and mitoses are prominent features. By immunohistochemistry the tumor cells express thyroglobulin and not calcitonin. Insular carcinoma is associated with a worse prognosis than well-differentiated thyroid carcinomas, but is significantly better than anaplastic thyroid carcinoma [331–333]. The extent of the poorly differentiated component in a well-differentiated thyroid tumor can affect the prognosis; tumors with >10% of the poorly differentiated component are associated with frequent regional recurrences, distant metastases, and poor prognosis [334].

### **10.7.2.3 Anaplastic Thyroid Tumors**

Anaplastic carcinomas are a group of high-grade thyroid carcinomas that are usually undifferentiated histologically and advertently have a lethal outcome [155,335]. Synonyms for anaplastic carcinoma include: undifferentiated, dedifferentiated, and sarcomatoid carcinoma. These tumors have represented approximately 10% of thyroid malignancies in older publications [155,336]. The tumor is more commonly seen in elderly females who present with a rapidly enlarging mass that often results in dyspnea. Risk factors are largely unknown but may include history of radiation and iodine deficiency [155]. A precursor well-differentiated thyroid carcinoma (papillary, follicular, or Hürthle cell) may be observed [337].



**Fig. 10.14** Anaplastic carcinoma. Pleomorphic spindle-shaped and epithelioid tumor cells

Grossly, the tumors are large with extensive intrathyroidal and extrathyroidal invasion. Surgical resection is often not performed because of the lesion's extent and diagnosis is commonly made on biopsy. Necrosis, vascular invasion, and mitoses are quite prominent [337]. Histologically, a variety of patterns have been described. The tumors are usually made up of a variety of cell types (Fig. 10.14). Most tumors are composed of giant cells and spindle cells although "squamous" differentiation is seen in about one third of cases [338]. Osteoclast-like giant cells are a common feature [339]. A "paucicellular" variant of anaplastic carcinoma has been described; it is characterized by dense fibrosis, calcification, and a poor patient outcome [340]. Spindle cell squamous anaplastic carcinoma may be the result of transformation of tall cell papillary carcinoma [257]. Carcinosarcoma of the thyroid has been described [341,342].

Electron microscopic and immunohistologic studies have indicated that almost all anaplastic thyroid tumors are indeed epithelial in nature [338,343]. By immunohistochemistry, anaplastic thyroid carcinomas should be positive for cytokeratin. Thyroglobulin immunostaining is often negative and thyroid transcription factor can be rarely positive in anaplastic carcinoma [343].

#### 10.7.2.4 Thyroid Sarcoma

Sarcomas of the thyroid are rare; fibrosarcomas, leiomyosarcomas, and angiosarcomas have been described [344,345]. Angiosarcoma of thyroid has been most commonly described from the mountainous regions of the world (Alpine regions of Europe, the Andes in South America, and the Himalayas in Asia)

[344,346]. Clinically, the affected patients resemble those with anaplastic carcinoma. By gross and histologic examination these tumors resemble angiosarcomas of soft tissue. These tumors generally lack the usual histologic features and exceptional aggressiveness of anaplastic carcinomas, but they are neither typical follicular nor papillary carcinomas.

#### 10.7.2.5 Squamous Cell Carcinoma, Mucoepidermoid Carcinoma, and Intrathyroidal Thymoma-like Neoplasms

*Squamous cell carcinoma* in thyroid occurs usually in association with papillary or anaplastic carcinoma [257]. Rarely, squamous cell carcinoma appears as an entity independent of any other form of thyroid cancer and behaves in an aggressive fashion with poor prognosis [347]. The major differential diagnosis is metastatic squamous carcinoma, especially from the head and neck, lungs, or esophagus.

*Mucoepidermoid carcinoma* is a distinctive variant of thyroid carcinoma. It is composed of solid masses of squamous cells and mucin-producing cells, sometimes forming glands [348]. Some authors consider that this lesion is a variant of papillary carcinoma; all cases show thyroglobulin expression [108,349]. The prognosis of thyroid mucoepidermoid carcinoma is quite good. Lesions may metastasize to regional nodes and rarely distantly. Death from disease is rare [108].

*Sclerosing mucoepidermoid carcinoma with eosinophilia* is usually seen in a background of lymphocytic thyroiditis and is characterized by tumor cells arranged in small sheets, anastomosing trabeculae, and narrow strands associated with dense fibrosis and numerous eosinophils. While these lesions may metastasize to lymph nodes and show extracapsular spread, vascular invasion, and perineural invasion, death due to disease is uncommon. The tumor cells stain negative for thyroglobulin and calcitonin and positive for cytokeratin [108,350,351].

There is no consensus regarding the origin of these tumors. Some studies have suggested that on the basis of immunoprofile both these tumors have different origins; mucoepidermoid carcinoma shows follicular derivation, and sclerosing mucoepidermoid carcinoma is derived from ultimobranchial body nests/solid cell nests [108].

Rare thyroid tumors composed of spindled epithelial cells arranged in nests, sometimes associated with mucous microcysts, and resembling thymomas (*spindled and epithelial tumor with thymus-like dif-*

*ferentiation; SETTLE*) have been reported [352,353]. Neoplasms resembling thymic carcinomas have also been described (*carcinoma with thymus-like differentiation; CASTLE*) in thyroid. These lesions may originate from branchial pouch remnants within and adjacent to the thyroid [352,354–356].

### 10.7.2.6 Medullary Carcinoma

Medullary thyroid carcinoma comprises less than 10% of all thyroid malignancies [357–361]. This tumor is of great diagnostic importance because of its aggressiveness, its close association with multiple endocrine neoplasia syndromes (MEN2A and 2B), and a relationship to a C cell hyperplasia, a probable precursor lesion [362]. While the majority of medullary carcinomas are sporadic, about 10–20% are familial [362]. Since these familial cases have been identified, a gene associated with medullary carcinoma has been identified on chromosome 10 and involves mutations in the RET oncogene [363–365].

Medullary carcinoma can affect patients of any age; most affected individuals are adults with an average age of about 50 years. However, in familial cases, children can be affected; also in these instances the age of diagnosis tends to be younger (mean age about 20 years) [361,366]. Although sporadic medullary carcinomas are seen more commonly in women, familial cases have a slight female to equal sex ratio, since an autosomal dominant mode of inheritance is present [367,368].

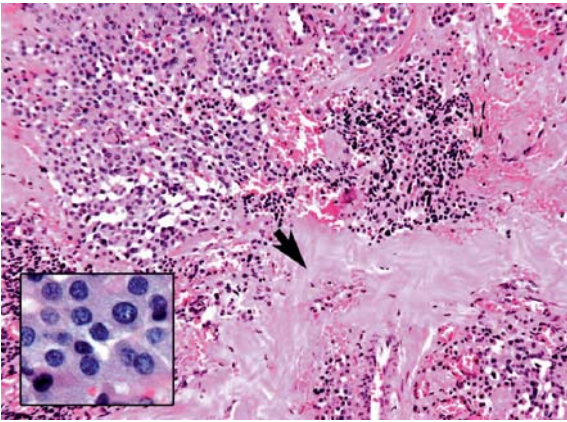
Clinically sporadic medullary carcinoma will present with a thyroid nodule that is painless but firm. In up to 50% of cases, obvious nodal metastases will be present at the time of diagnosis. Distant metastases, such as to lung, bone, or liver, may also be noted initially in about 15–25% of cases. When the tumor produces excess hormone other than calcitonin, the presenting symptoms may be related to that hormone hypersecretion [adrenocorticotrophic hormone (ACTH), prostaglandin] [369,370].

In the familial lesions there are associated endocrine and/or neuroendocrine lesions. Sipple's syndrome [multiple endocrine neoplasia (MEN) type 2A] [371] consists of medullary thyroid cancer and C cell hyperplasia, adrenal pheochromocytoma and adrenal medullary hyperplasia, and parathyroid hyperplasia [372]. Studies have shown that the gene responsible for familial medullary carcinoma is RET [373,374]; mutations in RET (different from the RET translocation in papillary carcinoma) are found in the tumors and the germline of patients with familial medullary

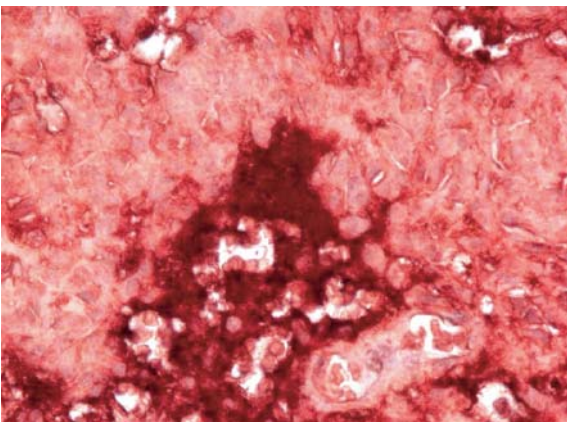
carcinomas and the MEN type 2 syndromes [373–375]. Mutations in specific codons have been correlated with clinical behavior and symptomatology in some families [374]. MEN type 2B consists of medullary thyroid carcinoma and C cell hyperplasia, pheochromocytoma and adrenal medullary hyperplasia, mucosal neuromas, gastrointestinal ganglioneuromas, and musculoskeletal abnormalities [376–379]. These patients may have familial disease (over 50% do), and some cases arise apparently as spontaneous mutations. These patients have biologically aggressive medullary carcinoma and may succumb to metastases at an early age. MEN2B shows similarity to von Recklinghausen's disease since in neurofibromatosis similar lesions are found in the gastrointestinal tract, and pheochromocytomas are common [378,379]. Nerve growth factor has been identified in some medullary carcinomas of these patients; it has been postulated that this product of the tumor may be responsible for the neural lesions seen in the MEN type 2B patients [380]. However, the neural lesions often precede by many years the development of medullary cancer. In MEN type 2B, the tumors and germline mutations in RET are found on codon 918—an intracellular focus of the RET oncogene [381,382].

Medullary carcinoma is usually located in the area of highest C cell concentration, i.e., the lateral upper two thirds of the gland. In familial cases, multiple small nodules may be detected grossly and, rarely, lesions may be found in the isthmus. The tumors range in size from barely visible to several centimeters. Many medullary carcinomas are grossly circumscribed but some will show infiltrative borders. The typical medullary carcinoma may be microscopically circumscribed or more likely will be infiltrating into the surrounding thyroid. The pattern of growth is of tumor cells arranged in nests separated by varying amounts of stroma. The tumor nests are composed of round, oval, or spindle-shaped cells; often there is isolated cellular pleomorphism or even multinucleated cells (Fig. 10.15) [383,384]. The tumor stroma characteristically contains amyloid although this is not necessary for the diagnosis as about 25% of medullary carcinomas do not contain amyloid (Fig. 10.15) [361,385,386]. The amyloid is most likely derived from procalcitonin and indeed immunohistochemical stains for calcitonin often stain the amyloid [37,386]. Calcifications in areas of amyloid deposition are characteristically present. The tumors commonly invade lymphatics and veins [384].

Several variants of medullary carcinoma on the basis of growth pattern have been described. These include: papillary variant, follicular variant, encap-



**Fig. 10.15** Medullary carcinoma. Tumor cells arranged in nests and round nuclei with finely granular chromatin (*inset*) in a background of stroma and amyloid (*arrow*)



**Fig. 10.16** Medullary carcinoma showing positive immunostaining with calcitonin antibody. The tumor is staining dark, and light staining is seen in the background amyloid

sulated variant, small cell variant, giant cell variant, oncocytic variant, and clear cell variant [389–391].

By immunohistochemistry, the majority of medullary carcinomas express low molecular weight cytokeratin, calcitonin (Fig. 10.16), calcitonin gene-related peptide, and thyroid transcription factor (TTF1). In addition, many tumors express CEA, which may also be elevated in the serum [392–394]. A variety of other peptides may be found in tumor cells including somatostatin, vasoactive intestinal peptide, and synaptophysin [395,396]. Some studies have also identified polysialic acid (neural cell adhesion molecule) in medullary carcinomas but not in other thyroid tumors [397].

Occasional lesions (and often these are small cell type) do not contain immunoreactive calcitonin. In order to accept a calcitonin-free tumor of the thyroid

as a medullary carcinoma, it should arise in a familial setting or occur in a thyroid with unequivocal C cell hyperplasia [398]. Immunoreactivity for calcitonin gene-related peptide would add proof to the histogenetic nature of such a lesion.

### **Prognostic Factors**

From the clinical standpoint, stage is the most important variable for prognosis [399–401]. A tumor confined to the thyroid without nodal or distant metastases is associated with prolonged survival. Several workers have found that younger patients (under age 40), especially women, fare somewhat better than the whole group of medullary cancer patients [399,402]. Patients who are discovered by screening because they are members of affected families often have very small tumors and can be cured by thyroidectomy. Patients with Sipple's syndrome tend to have less aggressive tumors than the sporadic group whereas the patients with MEN type 2B have aggressive lesions [401,403,404]. Pathologic features that have been related to prognosis include tumor pattern, amyloid content, pleomorphism, necrosis, mitotic activity, and DNA aneuploidy [405].

### **Mixed Follicular and Medullary Carcinoma**

These controversial tumors show thyroglobulin and calcitonin immunoreactivity and ultrastructural evidence of differentiation along two cell lines. Some of the series of these tumors may have been confusing, with trapping of follicles at the invading edge of the medullary carcinoma and diffusion of thyroglobulin into the medullary carcinoma; this may result in diagnosis of mixed tumors showing immunostaining for both hormones. Caution should be taken when making the diagnosis of mixed medullary and follicular-derived carcinomas [406–409].

### **Micromedullary Carcinoma**

A few medullary carcinomas are discovered incidental to thyroid operations for other conditions, at autopsy, or because of an elevated serum calcitonin. The so-called *micromedullary carcinomas* (equivalent to micropapillary carcinoma and defined as tumors of 1 cm or less) have an excellent prognosis if confined to the gland [410,411]. Some of the micromedullary cancers arise in the background of chronic thyroiditis

and may be associated with C cell hyperplasia even in the absence of familial disease [412]. Some of these patients have hypothyroidism and elevated TSH levels. Hence this type of C cell hyperplasia and micromedullary carcinoma may represent a secondary “reactive” phenomenon leading to early neoplastic change [410,412–414]. The non-tumoral parenchyma should be examined for evidence of C cell hyperplasia in a thyroid removed for a medullary carcinoma. Occasionally, the gland contains moderate to severe autoimmune thyroiditis, adenomatoid nodules, or another follicular-derived thyroid cancer [414–416].

### 10.7.3 Lymphoma

Secondary involvement of the thyroid by lymphoma has been reported in 20% of patients dying from generalized lymphoma. Primary lymphoma of the thyroid is uncommon but not rare. Most patients may have a history of diffuse goiter (probably the result of autoimmune thyroiditis) that has suddenly increased in size.

Most thyroid lymphomas are diffuse in type. Virtually all examples are B cell types; many may be extranodal lymphomas that arise in mucosa-associated lymphoid tissue (MALT) especially in the gastrointestinal tract. Some patients have typical plasmacytomas and these have a good prognosis. Hodgkin’s disease is extremely rare. Malignant lymphoma should be differentiated from advanced autoimmune thyroiditis; this distinction requires assessment of lymphocyte clonality by special studies (e.g., flow cytometry, gene rearrangement) [106,107,417–419].

### 10.7.4 Thyroid Tumors in Unusual Locations

Although clinically significant *lingual thyroid* is an unusual disorder, and microscopic remnants of thyroid tissue have been described in 9.8% of tongues examined at autopsy. Rare cases of thyroid carcinoma arising in lingual thyroid are recorded [420].

Neoplasms arising in association with the *thyroglossal duct* might be expected to be squamous carcinomas, but these are extremely rare; indeed, most tumors occurring in this setting have been thyroid carcinomas and most are described as papillary. Medullary carcinoma has not been described; since the parafollicular cells are not found in the median thyroid, this is not unexpected. The clinical presentation of thyroglossal duct carcinoma is identical to that of benign thyroglossal duct cysts, i.e., a swelling in the anterior neck [27,421,422].

When the diagnosis of thyroglossal cyst-associated thyroid cancer is made, the question of its origin arises. Does this tumor represent a metastasis from a primary lesion in the gland, or is the primary site in the region of the gland, or is the primary site in the region of the cyst? In rare cases in which the thyroid was examined pathologically, areas of papillary carcinoma were found in the gland [30,423]. Most authors studying this problem conclude that the thyroglossal carcinoma is a primary tumor arising in remnants of thyroid associated with the duct; in those few cases where intrathyroidal tumor has been found, this was considered a separate primary [30,421] although molecular analyses have not yet been reported to settle this question.

Malignant tumors arising in thyroid tissue located within the trachea or larynx are very rare, but have been reported [424].

Carcinomas, usually papillary subtype, and lesions that resemble carcinoid tumors can arise in struma ovarii [425–427].

### 10.7.5 Metastatic Neoplasms

Tumors metastasize to the thyroid via direct extension from tumors in adjacent structures, by retrograde lymphatic spread, or hematogenously. Carcinomas of the larynx, pharynx, trachea, and esophagus can invade the thyroid directly. In these cases the distinction from a thyroid primary is usually not difficult. Retrograde extension via lymphatic routes into the thyroid is unusual. In theory, at least, any tumor involving cervical lymph nodes could extend into the thyroid by this mechanism. Hematogenous metastases to the thyroid vary according to tumor type [329]. Carcinomas of the kidney, lung, and colon and melanoma are most commonly found [329]. Such lesions are often solitary, circumscribed masses; they may appear quite compatible with a primary tumor. Resemblance to colonic adenocarcinoma, breast cancer, or pigmented melanoma reassures that this is a metastasis. However, clear cell carcinoma of the kidney, as noted above, may present a problem [329,428–430].

## 10.8 Frozen Section Diagnosis and the Thyroid

Before the advent of fine- and large-needle biopsy, the method most often used in diagnosis of thyroid nodules was intraoperative frozen section. The nodule or preferably the thyroid lobe was excised and a representative portion (preferably encompassing nodule

capsule–thyroid interface) was prepared for frozen section and interpretation by a pathologist. In those cases in which the diagnosis of papillary, medullary, or anaplastic cancer was given, appropriate surgery was immediately undertaken.

Even with frozen section, however, despite recommendations of sampling two or even four different areas, the diagnosis of follicular carcinomas was notoriously difficult. In many cases, the diagnosis rendered is “follicular lesion diagnosis deferred to permanent sections” [299,300,431].

Several studies have evaluated frozen section and fine-needle aspirate (FNA) diagnostic results for thyroid nodules [432–434]. Although frozen section diagnosis may be specific (90–97%), it is not sensitive (60%). In addition, deferred diagnoses at frozen section do nothing to alter the operative procedure or guide the surgeon [299]. Frozen section results influenced the surgical approach in only a small percentage of cases. Also, in the era of cost containment, it does not seem justified to perform frozen sections for the intraoperative diagnosis of thyroid nodules; the initial approach to a thyroid nodule should be an aspiration biopsy (FNA) [299,435,436]. If the diagnosis rendered on FNA is definitely malignant, the surgeon should proceed with the appropriate surgery for that malignant diagnosis. If the FNA diagnosis is suspicious for malignancy, and that the suspected lesion is papillary carcinoma or a variant thereof, intraoperative frozen section may be useful since the diagnosis relies on the nuclear morphology and not the finding of invasion. If the FNA diagnosis is “neoplasm” and therefore non-committal as to the type, frozen section will not provide a definitive diagnosis and therefore should not be requested [37,248,251,437].

## References

1. Dozois RR, Beahrs OH (1977) Surgical anatomy and technique of thyroid and parathyroid surgery. *Surg Clin North Am* 57:647–661
2. Akimova RN, Zotikov LA (1969) [An electron microscope study of thyroid gland cells under normal conditions and during the carcinogenic process in golden hamsters]. *Vopr Onkol* 15:68–75
3. Mansberger AR, Jr., Wei JP (1993) Surgical embryology and anatomy of the thyroid and parathyroid glands. *Surg Clin North Am* 73:727–746
4. Miller FR (2003) Surgical anatomy of the thyroid and parathyroid glands. *Otolaryngol Clin North Am* 36:1–7, vii
5. Zampi G, Bianchi S, Amorosi A, Vezzosi V (1994) [Thyroid cancer: anatomy and pathologic histology]. *Chir Ital* 46:4–7
6. Kondalenko VF, Kalinin AP, Odinkova VA (1970) [Ultrastructure of the normal and pathologic human thyroid gland]. *Arkh Patol* 32:25–34
7. Nesland JM, Sobrinho-Simoes M, Johannessen JV (1987) Scanning electron microscopy of the human thyroid gland and its disorders. *Scanning Microsc* 1:1797–1810
8. Sugiyama S (1971) The embryology of the human thyroid gland including ultimobranchial body and others related. *Ergeb Anat Entwicklungsgesch* 44:3–111
9. Kovalenko AE (1999) [Contemporary concepts of embryology and surgical anatomy of the thyroid gland]. *Klin Khir* 1999:38–42
10. Gibson W, Peng T, Croker B (1980) C-cell nodules in adult human thyroid: a common autopsy finding. *Am J Clin Pathol* 73:347–351
11. Wolfe H, DeLellis R, Voelkel E, Tashjian A (1975) Distribution of calcitonin containing cells in the normal neonatal human thyroid gland: a correlation of morphology with peptide content. *J Clin Endocrinol Metab* 41:1076–1081
12. Baschieri L, Castagna M, Fierabracci A, Antonelli A, Del Guerra P, Squartini F (1989) Distribution of calcitonin- and somatostatin-containing cells in thyroid lymphoma and in Hashimoto's thyroiditis. *Appl Pathol* 7:99–104
13. Dhillon AP, Rode J, Leatham A, Papadaki L (1982) Somatostatin: a paracrine contribution to hypothyroidism in Hashimoto's thyroiditis. *J Clin Pathol* 35:764–770
14. O'Toole K, Fenoglio-Preiser C, Pushparaj N (1985) Endocrine changes associated with the human aging process. III. Effect of age on the number of calcitonin immunoreactive cells in the thyroid gland. *Hum Pathol* 16:991–1000
15. Guyétant S, Rousselet MC, Durigon M, et al (1997) Sex-related C cell hyperplasia in the normal human thyroid: a quantitative autopsy study. *J Clin Endocrinol Metab* 82:42–47
16. Harach H (1986) Solid cell nests of the human thyroid in early stages of postnatal life. *Acta Anat (Basel)* 127:262–264
17. Harach H (1988) Solid cell nests of the thyroid. *J Pathol* 155:191–200
18. Harach HR (1987) Mixed follicles of the human thyroid gland. *Acta Anat (Basel)* 129:27–30
19. Baloch ZW, LiVolsi VA (1999) Oncocytic lesions of the neuroendocrine system. *Semin Diagn Pathol* 16:190–199
20. Weiss ML, Deckart H, Pilz R, Deckart E, Kleinau E (1984) [Oncocytes in thyroid gland aspirates. Differential diagnostic problem: tumor/thyroiditis]. *Radiobiol Radiother (Berl)* 25:765–768
21. Mikhailov IG, Vasil'ev NB, Smirnova EA (1980) [Comparative quantitative electron-microscopic study of the nucleoli of human thyroid oncocytes and follicular cells]. *Arkh Patol* 42:32–36
22. Mitchell JD, Kirkham N, Machin D (1984) Focal lymphocytic thyroiditis in Southampton. *J Pathol* 144:269–273

23. Kabel PJ, Voorbij HA, van der Gaag RD, Wiersinga WM, de Haan M, Drexhage HA (1987) Dendritic C-cells in autoimmune thyroid disease. *Acta Endocrinol Suppl (Copenh)* 281:42–48
24. Nakamura Y, Watanabe M, Matsuzuka F, Maruoka H, Miyauchi A, Iwatani Y (2004) Intrathyroidal CD4+ T lymphocytes express high levels of Fas and CD4+ CD8+ macrophages/dendritic C-cells express Fas ligand in autoimmune thyroid disease. *Thyroid* 14:819–824
25. Katz AD, Hachigian M (1988) Thyroglossal duct cysts. A thirty year experience with emphasis on occurrence in older patients. *Am J Surg* 155:741–744
26. Sturgis EM, Miller RH (1988) Thyroglossal duct cysts. *J La State Med Soc* 145:459–461
27. Topf P, Fried MP, Strome M (1988) Vagaries of thyroglossal duct cysts. *Laryngoscope* 98:740–742
28. Allard R (1982) The thyroglossal cyst. *Head Neck Surg* 5:134–140
29. Baughman R (1972) Lingual thyroid and lingual thyroglossal tract remnants. *Oral Surg Oral Med Oral Pathol* 34:781–798
30. Heshmati HM, Fatourechi V, van Heerden JA, Hay ID, Goellner JR (1997) Thyroglossal duct carcinoma: report of 12 cases. *Mayo Clin Proc* 72:315–319
31. Apel RL, Asa SL, Chalvardjian A, LiVolsi VA (1994) Intrathyroidal lymphoepithelial cysts of probable branchial origin [see comments]. *Hum Pathol* 25:1238–1242
32. Carpenter GR, Emery JL (1976) Inclusions in the human thyroid. *J Anat* 122:77–89
33. Harach HR, Vujanic GM, Jasani B (1993) Ultimobranchial body nests in human fetal thyroid: an autopsy, histological, and immunohistochemical study in relation to solid cell nests and mucoepidermoid carcinoma of the thyroid. *J Pathol* 169:465–469
34. LiVolsi VA (1990) Surgical pathology of the thyroid. Saunders, Philadelphia
35. Rosai J, Carcangui ML, DeLellis RA (1992) Tumors of the thyroid gland, vol 3rd series, fascicle 5. Armed Forces Institute of Pathology, Washington, DC
36. Hathaway BM (1965) Innocuous accessory thyroid nodules. *Arch Surg* 90:222–227
37. Baloch Z, LiVolsi VA (2002) Pathology of the thyroid gland. Churchill Livingstone, Philadelphia
38. Gerard-Marchant R (1964) Thyroid follicle inclusions in cervical lymph nodes. *Arch Pathol Lab Med* 77:637–643
39. Roth L (1965) Inclusions of nonneoplastic thyroid tissue within cervical lymph nodes. *Cancer* 18:105–111
40. Meyer J, Steinberg L (1969) Microscopically benign thyroid follicles in cervical lymph nodes. *Cancer* 24:301–311
41. Bataskis J, Nishiyama R, Schmidt R (1963) “Sporadic goiter syndrome”: a clinicopathologic analysis. *Am J Clin Pathol* 30:241–251
42. Johnson J (1949) Adenomatous goiters with and without hyperthyroidism. *Arch Surg* 59:1088–1099
43. Struder H, Ramelli F (1982) Simple goiter and its variants: euthyroid and hyperthyroid. *Endocr Rev* 3:40–61
44. Weaver D, Batsakis J, Nishiyama R (1969) Relationship of iodine to “lymphocytic goiter.” *Arch Surg* 98:183–185
45. Brown R, Jackson I, Pohl S, Reichlin S (1978) Do thyroid stimulating immunoglobulins cause nontoxic and toxic multinodular goiter? *Lancet* 1:904–906
46. Gaitan E, Nelson NC, Poole GV (1991) Endemic goiter and endemic thyroid disorders. *World J Surg* 15:205–215
47. Braverman LE (2001) The physiology and pathophysiology of iodine and the thyroid. *Thyroid* 11:405
48. Ghossein RA, Rosai J, Heffess C (1997) Dyshormonogenetic goiter: a clinicopathologic study of 56 cases. *Endocr Pathol* 8:283–292
49. Rosenthal D, Carvalho-Guimaraes DP, Knobel M, Medeiros-Neto GA (1990) Dyshormonogenetic goiter: presence of an inhibitor of normal human thyroid peroxidase. *J Endocrinol Invest* 13:901–904
50. Bala TS, Janardanasarma MK, Raghunath M (1996) Dietary goitrogen-induced changes in the transport of 2-deoxy-d-glucose and amino acids across the rat blood-brain barrier. *Int J Dev Neurosci* 14:575–583
51. Fenwick GR, Griffiths NM (1981) The identification of the goitrogen (-)-5-vinylloxazolidine-2-thione (goitroin) as a bitter principle of cooked Brussels sprouts (*Brassica oleracea* L. var. gemmifera). *Z Lebensm Unters Forsch* 172:90–92
52. Gabrilove JL, Dorrance WR, Soffer LJ (1952) Effect of corticotropin, cortisone and desoxycorticosterone on thyroid weight of the goitrogen-treated rat. *Am J Physiol* 169:565–567
53. Amdisen A, Jensen SE, Olsen T, Schou M (1968) [Development of goiter during lithium treatment]. *Ugeskr Laeger* 130:1515–1518
54. Maloof F, Wang CA, Vickery AL, Jr (1975) Nontoxic goiter-diffuse or nodular. *Med Clin North Am* 59:1221–1232
55. Struder H, Peter H, Gerber H (1987) Morphologic and functional changes in developing goiters. In: Hall R, Koberling J (eds) *Thyroid disorders associated with iodine deficiency and excess*. Raven, New York
56. Murray D (1998) *The thyroid gland*. Blackwell Science, Malden, MA
57. Barsano C, DeGroot L (1979) Dyshormonogenetic goiter. *Baillieres Clin Endocrinol Metab* 8:145–165
58. Medeiros-Neto G, Bunduki V, Tomimori E, et al (1997) Prenatal diagnosis and treatment of dyshormonogenetic fetal goiter due to defective thyroglobulin synthesis. *J Clin Endocrinol Metab* 82:4239–4242
59. Ramelli F, Studer H, Bruggisser D (1982) Pathogenesis of thyroid nodules in multinodular goiter. *Am J Pathol* 109:215–223
60. Fialho NJ, de Oliveira CA (1971) [Colloid goiter (observations on 100 operated and treated cases)]. *Rev Bras Med* 28:314–326
61. Greer MA, Studer H, Kendall JW (1967) Studies on the pathogenesis of colloid goiter. *Endocrinology* 81:623–632



62. Nair K (1951) Adenoma of thyroid: simple adenomatous goiter. *Antiseptic* 48:716–724
63. Hirose H, Noguchi M, Sakata N, Tanaka S, Miyazaki I (1983) [Adenoma or adenomatous goiter with the clinical symptoms of hyperthyroidism]. *Horumon To Rinsho* 31(suppl):95–98
64. Apel RL, Ezzat S, Bapat BV, Pan N, LiVolsi VA, Asa SL (1995) Clonality of thyroid nodules in sporadic goiter. *Diagn Mol Pathol* 4:113–121
65. Hicks DG, LiVolsi VA, Neidich JA, Puck JM, Kant JA (1990) Clonal analysis of solitary follicular nodules in the thyroid. *Am J Pathol* 137:553–562
66. Miller JM (1975) Plummer's disease. *Med Clin North Am* 59:1203–1216
67. LiVolsi VA (1994) The pathology of autoimmune thyroid disease: a review. *Thyroid* 4:333–339
68. Margolick JB, Hsu SM, Volkman DJ, Burman KD, Fauci AS (1984) Immunohistochemical characterization of intrathyroid lymphocytes in Graves' disease. Interstitial and intraepithelial populations. *Am J Med* 76:815–821
69. Misaki T, Konishi J, Nakashima T, et al (1985) Immunohistological phenotyping of thyroid infiltrating lymphocytes in Graves' disease and Hashimoto's thyroiditis. *Clin Exp Immunol* 60:104–110
70. Totterman TH (1978) Distribution of T-, B-, and thyroglobulin-binding lymphocytes infiltrating the gland in Graves' disease, Hashimoto's thyroiditis, and de Quervain's thyroiditis. *Clin Immunol Immunopathol* 10:270–277
71. Misaki T, Konishi J, Arai K, et al (1987) HLA-DR antigen expression on intrathyroidal lymphocytes and thyrocytes in Hashimoto's thyroiditis and Graves' disease: an immunohistological study. *Endocrinol Jpn* 34:257–262
72. Weetman AP, Gunn C, Hall R, McGregor AM (1985) Thyroid autoantigen-induced lymphocyte proliferation in Graves' disease and Hashimoto's thyroiditis. *J Clin Lab Immunol* 17:1–6
73. Brinkane A, Ounadi-Corbille W, Bellamy J, Leroy-Terquem E (2004) [Hyperplasia of the thymus in Graves' disease. A case report]. *Rev Pneumol Clin* 60:239–241
74. Kawai K, Tamai H, Mori T, et al (1993) Thyroid histology of hyperthyroid Graves' disease with undetectable thyrotropin receptor antibodies. *J Clin Endocrinol Metab* 77:716–719
75. Hirota Y, Tamai H, Hayashi Y, et al (1986) Thyroid function and histology in forty-five patients with hyperthyroid Graves' disease in clinical remission more than ten years after thionamide drug treatment. *J Clin Endocrinol Metab* 62:165–169
76. Cassano C (1971) [Dyshormonogenetic goiter caused by altered synthesis of thyroglobulin]. *Recenti Prog Med* 50:9–23
77. Vickery AL, Jr (1981) The diagnosis of malignancy in dyshormonogenetic goiter. *Clin Endocrinol Metab* 10:317–335
78. Roti E, Uberti ED (2001) Iodine excess and hyperthyroidism. *Thyroid* 11:493–500
79. Strauss A, Trujillo M (1986) Lithium-induced goiter and voice changes. *J Clin Psychopharmacol* 6:120–121
80. Mizukami Y, Funaki N, Hashimoto T, Kawato M, Michigishi T, Matsubara F (1988) Histologic features of thyroid gland in a patient with bromide-induced hypothyroidism. *Am J Clin Pathol* 89:802–805
81. Imai C, Kakihara T, Watanabe A, et al (2002) Acute suppurative thyroiditis as a rare complication of aggressive chemotherapy in children with acute myelogenous leukemia. *Pediatr Hematol Oncol* 19:247–253
82. Lambert MJ 3rd, Johns ME, Mentzer R (1980) Acute suppurative thyroiditis. *Am Surg* 46:461–463
83. Golshan MM, McHenry CR, de Vente J, Kalajyan RC, Hsu RM, Tomashefski JF (1997) Acute suppurative thyroiditis and necrosis of the thyroid gland: a rare endocrine manifestation of acquired immunodeficiency syndrome. *Surgery* 121:593–596
84. Leesen E, Janssen L, Smet M, Breysem L (2001) Acute suppurative thyroiditis. *JBR-BTR* 84:68
85. Hnilica P, Nyulassy S (1985) Plasma cells in aspirates of goitre and overt permanent hypothyroidism following subacute thyroiditis. Preliminary report. *Endocrinol Exp* 19:221–226
86. Weetman AP, Smallridge RC, Nutman TB, Burman KD (1987) Persistent thyroid autoimmunity after subacute thyroiditis. *J Clin Lab Immunol* 23:1–6
87. Greene J. Subacute thyroiditis (1971) *Am J Med* 51:97–108
88. Bastenie P, Bonnyns M, Neve P (1972) Subacute and chronic granulomatous thyroiditis. In: Bastenie PA, Ermans AM (eds) *Thyroiditis and thyroid function, clinical, morphological and physiological studies*. Pergamon, Oxford, pp 69–97
89. Cordray JP, Nys P, Merceron RE, Augusti A (2001) [Frequency of hypothyroidism after de Quervain thyroiditis and contribution of ultrasonographic thyroid volume measurement]. *Ann Med Interne (Paris)* 152:84–88
90. Harach HR, Williams ED (1990) The pathology of granulomatous diseases of the thyroid gland. *Sarcoidosis* 7:19–27
91. Meachim G, Young M (1963) De Quervain's subacute granulomatous thyroiditis: histological identification and incidence. *J Clin Pathol* 16:189–199
92. Carney JA, Moore SB, Northcutt RC, Woolner LB, Stillwell GK (1975) Palpation thyroiditis (multifocal granulomatous folliculitis). *Am J Clin Pathol* 64:639–647
93. Harach HR (1993) Palpation thyroiditis resembling C cell hyperplasia. Usefulness of immunohistochemistry in their differential diagnosis. *Pathol Res Pract* 189:488–490
94. Mizukami Y, Michigishi T, Kawato M, et al (1982) Chronic thyroiditis: thyroid function and histologic correlations in 601 cases. *Hum Pathol* 23:980

95. Hayashi Y, Tamai H, Fukata S, et al (1985) A long term clinical, immunological, and histological follow-up study of patients with goitrous chronic lymphocytic thyroiditis. *J Clin Endocrinol Metab* 61:1172–1178
96. Roitt IM, De Carvalho LC (1982) The immunological basis of autoimmune disease. *Ciba Found Symp* 1982:22–34
97. Podleski WK (1971) Quantitative distribution of IgG, IgM and IgA immunoglobulins in lymphocytic thyroiditis of the Hashimoto type. *Arch Immunol Ther Exp* 19:431–438
98. Tomer Y, Ban Y, Concepcion E, et al (2003) Common and unique susceptibility loci in Graves and Hashimoto diseases: results of whole-genome screening in a data set of 102 multiplex families. *Am J Hum Genet* 73:736–747
99. Fatourechi V, McConahey WM, Woolner LB (1971) Hyperthyroidism associated with histologic Hashimoto's thyroiditis. *Mayo Clin Proc* 46:682–689
100. Mizukami Y, Michigishi T, Hashimoto T, et al (1988) Silent thyroiditis: a histologic and immunohistochemical study. *Hum Pathol* 19:423–431
101. Aozasa M, Amino N, Iwatani Y, et al (1989) Intrathyroidal HLA-DR-positive lymphocytes in Hashimoto's disease: increases in CD8 and Leu7 cells. *Clin Immunol Immunopathol* 52:516–522
102. Iwatani Y, Amino N, Mori H, et al (1983) T lymphocyte subsets in autoimmune thyroid diseases and subacute thyroiditis detected with monoclonal antibodies. *J Clin Endocrinol Metab* 56:251–254
103. Iwatani Y, Hidaka Y, Matsuzuka F, Kuma K, Amino N (1993) Intrathyroidal lymphocyte subsets, including unusual CD4+ CD8+ cells and CD3loTCR alpha beta lo/CD4-CD8 cells, in autoimmune thyroid disease. *Clin Exp Immunol* 93:430–436
104. Weetman AP, Volkman DJ, Burman KD, et al (1986) The production and characterization of thyroid-derived T-cell lines in Graves' disease and Hashimoto's thyroiditis. *Clin Immunol Immunopathol* 39:139–150
105. McIntosh RS, Watson PE, Weetman AP (1997) Analysis of the T cell receptor V alpha repertoire in Hashimoto's thyroiditis: evidence for the restricted accumulation of CD8+ T cells in the absence of CD4+ T cell restriction. *J Clin Endocrinol Metab* 82:1140–1146
106. Kossel P, Livolsi V (1999) Lymphoid lesions of the thyroid: review in light of the revised European-American lymphoma classification and upcoming World Health Organization classification. *Thyroid* 9:1273–1280
107. Lam KY, Lo CY, Kwong DL, Lee J, Srivastava G (1999) Malignant lymphoma of the thyroid. A 30-year clinicopathologic experience and an evaluation of the presence of Epstein-Barr virus. *Am J Clin Pathol* 112:263–270
108. Baloch ZW, Solomon AC, LiVolsi VA (2000) Primary mucoepidermoid carcinoma and sclerosing mucoepidermoid carcinoma with eosinophilia of the thyroid gland: a report of nine cases. *Mod Pathol* 13:802–807
109. Katz SM, Vickery AL Jr (1974) The fibrous variant of Hashimoto's thyroiditis. *Hum Pathol* 5:161–170
110. Papi G, Corrado S, Carapezzi C, De Gaetani C, Carani C (2003) Riedel's thyroiditis and fibrous variant of Hashimoto's thyroiditis: a clinicopathological and immunohistochemical study. *J Endocrinol Invest* 26:444–449
111. LiVolsi VA (1993) Postpartum thyroiditis. The pathology slowly unravels. *Am J Clin Pathol* 100:193–195
112. Weetman AP, Fung HY, Richards CJ, McGregor AM (1990) IgG subclass distribution and relative functional affinity of thyroid microsomal antibodies in postpartum thyroiditis. *Eur J Clin Invest* 20:133–136
113. Papi G, Corrado S, Carapezzi C, Corsello SM (2003) Postpartum thyroiditis presenting as a cold nodule and evolving to Graves' disease. *Int J Clin Pract* 57:556–558
114. Williams ED, Doniach I (1962) The post-mortem incidence of focal thyroiditis. *J Pathol Bacteriol* 83:255–264
115. Weaver DR, Deodhar SD, Hazard JB (1966) A characterization of focal lymphocytic thyroiditis. *Cleve Clin Q* 33:59–72
116. Vollenweider R, Stolkin I, Hedinger C (1982) [Focal lymphocytic thyroiditis and iodized salt prophylaxis. Comparative studies on goiter specimens at the Institute of Pathology of Zurich University]. *Schweiz Med Wochenschr* 112:482–488
117. Katsikas D, Shorthouse A, Taylor S (1976) Riedel's thyroiditis. *Br J Surg* 63:929–931
118. Arnott E, Greaves D (1965) Orbital involvement in Riedel's thyroiditis. *Br J Ophthalmol* 49:1–5
119. Bartholomew L, Cain J, Woolner L, Utz D, Ferris D (1963) Sclerosing cholangitis. Its possible association with Riedel's struma and fibrous retroperitonitis. *N Engl J Med* 269:8–12
120. Davies D, Furness P (1984) Riedel's thyroiditis with multiple organ fibrosis. *Thorax* 39:959–960
121. Rao C, Ferguson G, Kyle V (1973) Retroperitoneal fibrosis associated with Riedel's struma. *Can Med Assoc J* 108:1019–1021
122. Schwaegerle S, Bauer T, Esselstyn C (1988) Riedel's thyroiditis. *Am J Clin Pathol* 90:715–722
123. Papi G, Corrado S, Cesinaro AM, Novelli L, Smerieri A, Carapezzi C (2002) Riedel's thyroiditis: clinical, pathological and imaging features. *Int J Clin Pract* 56:65–67
124. Casoli P, Tumiatì B (1999) Hypoparathyroidism secondary to Riedel's thyroiditis. A case report and a review of the literature. *Ann Ital Med Int* 14:54–57
125. Meij S, Hausman R (1978) Occlusive phlebitis, a diagnostic feature in Riedel's thyroiditis. *Virchows Arch [A]*. 377:339–349
126. Harach HR, Williams ED (1983) Fibrous thyroiditis: an immunopathological study. *Histopathology* 7:739–751
127. Baloch ZW, Feldman MD, LiVolsi VA (2000) Combined Riedel's disease and fibrosing Hashimoto's thyroiditis: a report of three cases with two showing coexisting papillary carcinoma. *Endocr Pathol* 11:157–163

128. Bogazzi F, Bartalena L, Gasperi M, Braverman LE, Martino E (2001) The various effects of amiodarone on thyroid function. *Thyroid* 11:511–519
129. Smyrk T, Goellner J, Brennan M, Carney J (1987) Pathology of the thyroid in amiodarone associated thyrotoxicosis. *Am J Surg Pathol* 11:197–204
130. Cetta F, Montalto G, Petracchi M, Fusco A (1997) Thyroid cancer and the Chernobyl accident. Are long-term and long-distance side effects of fall-out radiation greater than estimated? *J Clin Endocrinol Metab* 82:2015–2017
131. Carr RF, LiVolsi VA (1989) Morphologic changes in the thyroid after irradiation for Hodgkin's and non-Hodgkin's lymphoma. *Cancer* 64:825–829
132. Aizawa T, Watanabe T, Suzuki N, et al (1998) Radiation-induced painless thyrotoxic thyroiditis followed by hypothyroidism: a case report and literature review. *Thyroid* 8:273–275
133. Avetisian IL, Gulchiv NV, Demidiuk AP, Stashuk AV (1996) Thyroid pathology in residents of the Kiev region, Ukraine, during pre- and post-Chernobyl periods. *J Environ Pathol Toxicol Oncol* 15:233–237
134. Nishiyama K, Kozuka T, Higashihara T, Miyauchi K, Okagawa K (1996) Acute radiation thyroiditis. *Int J Radiat Oncol Biol Phys* 36:1221–1224
135. Lindsay S, Dailey M, Jones M (1954) Histologic effects of various types of ionizing radiation on normal and hyperplastic human thyroid glands. *J Clin Endocrinol Metab* 14:1179–1219
136. Goldsmith JD, Lai ML, Daniele GM, Tomaszewski JE, LiVolsi VA (2000) Amyloid goiter: report of two cases and review of the literature. *Endocr Pract* 6:318–323
137. Hamed G, Heffess CS, Shmookler BM, Wenig BM (1995) Amyloid goiter. A clinicopathologic study of 14 cases and review of the literature. *Am J Clin Pathol* 104:306–312
138. Pastoloro GC, Asa SL (1994) Drug-related pigmentation of the thyroid associated with papillary carcinoma. *Arch Pathol Lab Med* 118:79–83
139. Gordon G, Sparano B, Kramer A, Kelly R, Latropoulos M (1984) Thyroid gland pigmentation and minocycline therapy. *Am J Pathol* 117:98–109
140. LiVolsi VA, Feind CR (1976) Parathyroid adenoma and nonmedullary thyroid carcinoma. *Cancer* 38:1391–1393
141. Mai KT, Landry DC, Thomas J, et al (2001) Follicular adenoma with papillary architecture: a lesion mimicking papillary thyroid carcinoma. *Histopathology* 39:25–32
142. LiVolsi VA (1996) Well differentiated thyroid carcinoma. *Clin Oncol (R Coll Radiol)* 8:281–288
143. LiVolsi VA, Baloch ZW (2004) Follicular neoplasms of the thyroid: view, biases, and experiences. *Adv Anat Pathol* 11:279–287
144. LiVolsi VA, Merino MJ (1994) Worrisome histologic alterations following fine-needle aspiration of the thyroid (WHAFFT). *Pathol Annu* 29:99–120
145. Oyama T, Vickery AL Jr, Preffer FI, Colvin RB (1994) A comparative study of flow cytometry and histopathologic findings in thyroid follicular carcinomas and adenomas. *Hum Pathol* 25:271–275
146. Harlow SP, Duda RB, Bauer KD (1992) Diagnostic utility of DNA content flow cytometry in follicular neoplasms of the thyroid. *J Surg Oncol* 50:1–6
147. Carney JA, Ryan J, Goellner JR (1987) Hyalinizing trabecular adenoma of the thyroid gland. *Am J Surg Pathol* 11:583–591
148. Chetty R, Beydoun R, LiVolsi VA (1994) Paraganglioma-like (hyalinizing trabecular) adenoma of the thyroid revisited. *Pathology* 26:429–431
149. LiVolsi VA (2000) Hyalinizing trabecular tumor of the thyroid: adenoma, carcinoma, or neoplasm of uncertain malignant potential? *Am J Surg Pathol* 24:1683–1684
150. Fonseca E, Nesland J, Sobrinho-Simoes M (1997) Expression of stratified epithelial type cytokeratins in hyalinizing trabecular adenoma supports their relationship with papillary carcinoma of the thyroid. *Histopathology* 31:330–335
151. Papotti M, Volante M, Giuliano A, et al (2000) RET/PTC activation in hyalinizing trabecular tumors of the thyroid. *Am J Surg Pathol* 24:1615–1621
152. Hazard JB, Kenyon R (1954) Atypical adenoma of the thyroid. *Arch Pathol* 58:554–563
153. Fukunaga M, Shinozaki N, Endo Y, Ushigome S (1992) Atypical adenoma of the thyroid. A clinicopathologic and flow cytometric DNA study in comparison with other follicular neoplasms. *Acta Pathol Jpn* 42:632–638
154. Lang W, Georgii A, Atay Z (1977) [Differential diagnosis between atypical adenoma and follicular carcinoma of the thyroid gland (author's translation)]. *Verh Dtsch Ges Pathol* 61:275–279
155. Carcangiu ML, Steeper T, Zampi G, Rosai J (1985) Anaplastic thyroid carcinoma. A study of 70 cases. *Am J Clin Pathol* 83:135–158
156. Carcangiu ML ZG, Pupi A, Castagnoli A, Rosai J (1985) Papillary carcinoma of the thyroid: a clinico-pathologic study of 241 cases treated at the University of Florence, Italy. *Cancer* 55:805–828
157. LiVolsi VA (1992) Papillary neoplasms of the thyroid. Pathologic and prognostic features. *Am J Clin Pathol* 97:426–434
158. Mazzaferri EYR (1981) Papillary thyroid carcinoma: a 10-year follow-up report of the impact of therapy in 576 patients. *Am J Med* 70:511–518
159. Furlan JC, Bedard YC, Rosen IB (2004) Clinicopathologic significance of histologic vascular invasion in papillary and follicular thyroid carcinomas. *J Am Coll Surg* 198:341–348
160. Mills SE, Allen MS Jr (1986) Congenital occult papillary carcinoma of the thyroid gland. *Hum Pathol* 17:1179–1181

161. Schottenfeld D, Gershman ST (1977) Epidemiology of thyroid cancer, part II. *Clin Bull* 7:98–104
162. Williams ED, Doniach I, Bjarnason O, Michie W (1977) Thyroid cancer in an iodide rich area: a histopathological study. *Cancer* 39:215–222
163. Harach HR, Escalante DA, Day ES (2002) Thyroid cancer and thyroiditis in Salta, Argentina: a 40-yr study in relation to iodine prophylaxis. *Endocr Pathol* 13:175–181
164. Petrova GV, Tereshchenko VP, Avetis'ian IL (1996) [The dynamics of thyroid diseases in the inhabitants of Kiev and Kiev Province after the accident at the Chernobyl Atomic Electric Power Station]. *Lik Sprava* 1996:67–70
165. Ron E, Kleinerman RA, Boice JD, LiVolsi VA, Flannery JT, Fraumeni JF (1987) A population-based case-control study of thyroid cancer. *J Natl Cancer Inst* 79:1–12
166. Pacini F, Vorontsova T, Molinaro E, et al (1999) Thyroid consequences of the Chernobyl nuclear accident. *Acta Paediatr Suppl* 88:23–27
167. Tamimi DM (2002) The association between chronic lymphocytic thyroiditis and thyroid tumors. *Int J Surg Pathol* 10:141–146
168. Nikiforova MN, Caudill CM, Biddinger P, Nikiforov YE (2002) Prevalence of RET/PTC rearrangements in Hashimoto's thyroiditis and papillary thyroid carcinomas. *Int J Surg Pathol* 10:15–22
169. Hunt JL, Baloch Z, Barnes EL, et al (2002) Loss of heterozygosity mutations of tumor suppressor genes in cytologically atypical areas of chronic lymphocytic thyroiditis. *Endocr Pathol* 13:23–30
170. La Vecchia C, Ron E, Franceschi S, et al (1999) A pooled analysis of case-control studies of thyroid cancer. III. Oral contraceptives, menopausal replacement therapy and other female hormones. *Cancer Causes Control* 10:157–166
171. Diaz NM, Mazoujian G, Wick MR (1991) Estrogen-receptor protein in thyroid neoplasms. An immunohistochemical analysis of papillary carcinoma, follicular carcinoma, and follicular adenoma. *Arch Pathol Lab Med* 115:1203–1207
172. Cetta F, Montalto G, Gori M, Curia MC, Cama A, Olschwang S (2000) Germline mutations of the APC gene in patients with familial adenomatous polyposis-associated thyroid carcinoma: results from a European cooperative study. *J Clin Endocrinol Metab* 85:286–292
173. Harach HR, Soubeyran I, Brown A, Bonneau D, Longy M (1999) Thyroid pathologic findings in patients with Cowden disease. *Ann Diagn Pathol* 3:331–340
174. Haggitt RC, Reid BJ (1986) Hereditary gastrointestinal polyposis syndromes. *Am J Surg Pathol* 10:871–887
175. Cetta F, Toti P, Petracci M, et al (1997) Thyroid carcinoma associated with familial adenomatous polyposis. *Histopathology* 31:231–236
176. Cetta F, Olschwang S, Petracci M, et al (1998) Genetic alterations in thyroid carcinoma associated with familial adenomatous polyposis: clinical implications and suggestions for early detection. *World J Surg* 22:1231–1236
177. Cetta F, Chiappetta G, Melillo RM, et al (1998) The ret/ptc1 oncogene is activated in familial adenomatous polyposis-associated thyroid papillary carcinomas. *J Clin Endocrinol Metab* 83:1003–1006
178. Dahia PL, Marsh DJ, Zheng Z, et al (1997) Somatic deletions and mutations in the Cowden disease gene, PTEN, in sporadic thyroid tumors. *Cancer Res* 57:4710–4713
179. Wirtzfeld DA, Petrelli NJ, Rodriguez-Bigas MA (2001) Hamartomatous polyposis syndromes: molecular genetics, neoplastic risk, and surveillance recommendations. *Ann Surg Oncol* 8:319–327
180. Meshikhes AW, Butt MS, Al-Saihati BA (2004) Combined parathyroid adenoma and an occult papillary carcinoma. *Saudi Med J* 25:1707–1710
181. Dralle H, Altenahr E (1979) Pituitary adenoma, primary parathyroid hyperplasia and papillary (non-medullary) thyroid carcinoma. A case of multiple endocrine neoplasia (MEN). *Virchows Arch A Pathol Anat Histol* 381:179–187
182. Francis IM, Das DK, Sheikh ZA, Sharma PN, Gupta SK (1995) Role of nuclear grooves in the diagnosis of papillary thyroid carcinoma. A quantitative assessment on fine needle aspiration smears. *Acta Cytol* 39:409–415
183. Riazmontazer N, Bedayat G (1991) Psammoma bodies in fine needle aspirates from thyroids containing nontoxic hyperplastic nodular goiters. *Acta Cytol* 35:563–566
184. Hunt JL, Barnes EL (2003) Non-tumor-associated psammoma bodies in the thyroid. *Am J Clin Pathol* 119:90–94
185. Hosoya T, Sakamoto A, Kasai N, Sakurai K (1983) [Nodal psammoma body in thyroid cancer as an indicator of cancer metastasis to the lymph node]. *Gan No Rinsho* 29:1336–1339
186. Mancini A, Rabitti C, Conte G, Gullotta G, De Marinis L (1993) [Lymphocytic infiltration in thyroid neoplasms. Preliminary prognostic assessments]. *Minerva Chir* 48:1283–1288
187. Gomez Saez JM, Gomez Arnaiz N, Sahun de la Vega M, Soler Ramon J (1997) [Prevalence and significance of lymphocyte infiltration in papillary carcinoma of the thyroid gland]. *Ann Med Interna* 14:403–405
188. Ruiz-Velasco R, Waisman J, Van Herle AJ (1978) Cystic papillary carcinoma of the thyroid gland. Diagnosis by needle aspiration with transmission electron microscopy. *Acta Cytol* 22:38–42
189. de los Santos ET, Keyhani-Rofagha S, Cunningham JJ, Mazzaferri EL (1990) Cystic thyroid nodules. The dilemma of malignant lesions. *Arch Intern Med* 150:1422–1427
190. Namba H, Matsuo K, Fagin JA (1990) Clonal composition of benign and malignant human thyroid tumors. *J Clin Invest* 86:120–125
191. Fusco A, Chiappetta G, Hui P, et al (2002) Assessment of RET/PTC oncogene activation and clonality in thyroid nodules with incomplete morphological evidence of papillary carcinoma: a search for the early precursors of papillary cancer. *Am J Pathol* 160:2157–2167

192. Hunt JL, LiVolsi VA, Baloch ZW, et al (2003) Microscopic papillary thyroid carcinoma compared with clinical carcinomas by loss of heterozygosity mutational profile. *Am J Surg Pathol* 27:159–166
193. Petkov R, Gavrailov M, Mikhailov I, Todorov G, Kutev N (1995) [Differentiated thyroid cancer: a study of the pathomorphological variants in 216 patients]. *Khirurgiia* 48:11–12
194. Akslen LA, LiVolsi VA (2000) Prognostic significance of histologic grading compared with subclassification of papillary thyroid carcinoma [see comments]. *Cancer* 88:1902–1908
195. Paessler M, Kreisel FH, LiVolsi VA, Akslen LA, Baloch ZW (2002) Can we rely on pathologic parameters to define conservative treatment of papillary thyroid carcinoma? *Int J Surg Pathol* 10:267–272
196. Santini L, Pezzullo L, D'Arco E, De Rosa N, Guerriero O, Salza C (1989) Lymph node metastases from an occult sclerosing carcinoma of the thyroid. A case report. *Ital J Surg Sci* 19:277–279
197. Akslen LA, LiVolsi VA (2000) Poorly differentiated thyroid carcinoma: it is important. *Am J Surg Pathol* 24:310–313
198. Tachikawa T, Kumazawa H, Kyomoto R, Yukawa H, Yamashita T, Nishikawa M (2001) [Clinical study on prognostic factors in thyroid carcinoma]. *Nippon Jibiinkoka Gakkai Kaiho* 104:157–164
199. Mazzaferri EL (1987) Papillary thyroid carcinoma: factors influencing prognosis and current therapy. *Semin Oncol* 14:315–332
200. Prasad ML, Pellegata NS, Huang Y, Nagaraja HN, Chappelle Ade L, Kloos RT (2005) Galectin-3, fibronectin-1, CITED-1, HBME1 and cytokeratin-19 immunohistochemistry is useful for the differential diagnosis of thyroid tumors. *Mod Pathol* 18:48–57
201. Casey MB, Lohse CM, Lloyd RV (2003) Distinction between papillary thyroid hyperplasia and papillary thyroid carcinoma by immunohistochemical staining for cytokeratin 19, galectin-3, and HBME-1. *Endocr Pathol* 14:55–60
202. Cheung CC, Ezzat S, Freeman JL, Rosen IB, Asa SL (2001) Immunohistochemical diagnosis of papillary thyroid carcinoma. *Mod Pathol* 14:338–342
203. Eimoto T, Naito H, Hamada S, Masuda M, Harada T, Kikuchi M (1987) Papillary carcinoma of the thyroid. A histochemical, immunohistochemical and ultrastructural study with special reference to the follicular variant. *Acta Pathol Jpn* 37:1563–1579
204. van Hoesven KH, Kovatich AJ, Miettinen M (1998) Immunocytochemical evaluation of HBME-1, CA 19-9, and CD-15 (Leu-M1) in fine-needle aspirates of thyroid nodules. *Diagn Cytopathol* 18:93–97
205. Baloch ZW, Abraham S, Roberts S, LiVolsi VA (1999) Differential expression of cytokeratins in follicular variant of papillary carcinoma: an immunohistochemical study and its diagnostic utility. *Hum Pathol* 30:1166–1171
206. Baloch ZW, LiVolsi VA (2002) The quest for a magic tumor marker: continuing saga in the diagnosis of the follicular lesions of thyroid. *Am J Clin Pathol* 118:165–166
207. Batistatou A, Zolota V, Scopa CD (2002) S-100 protein+ dendritic-cells and CD34+ dendritic interstitial cells in thyroid lesions. *Endocr Pathol* 13:111–115
208. Hiiasa Y, Nishioka H, Kitahori Y, et al (1991) Immunohistochemical detection of estrogen receptors in paraffin sections of human thyroid tissues. *Oncology* 48:421–424
209. Khan A, Baker SP, Patwardhan NA, Pullman JM (1998) CD57 (Leu-7) expression is helpful in diagnosis of the follicular variant of papillary thyroid carcinoma. *Virchows Arch* 432:427–432
210. Miettinen M, Karkkainen P (1996) Differential reactivity of HBME-1 and CD15 antibodies in benign and malignant thyroid tumours. Preferential reactivity with malignant tumours. *Virchows Arch* 429:213–219
211. Zedenius J, Auer G, Backdahl M, et al (1992) Follicular tumors of the thyroid gland: diagnosis, clinical aspects and nuclear DNA analysis. *World J Surg* 16:589–594
212. Grieco M, Santoro M, Berlingieri MT, et al (1990) PTC is a novel rearranged form of the ret proto-oncogene and is frequently detected in vivo in human thyroid papillary carcinomas. *Cell* 60:557–563
213. Santoro M, Rosati R, Grieco M, et al (1990) The ret proto-oncogene is consistently expressed in human pheochromocytomas and thyroid medullary carcinomas. *Oncogene* 5:1595–1598
214. Santoro M, Dathan NA, Berlingieri MT, et al (1994) Molecular characterization of RET/PTC3: a novel rearranged version of the RET proto-oncogene in a human thyroid papillary carcinoma. *Oncogene* 9:509–516
215. Inaba M, Umemura S, Satoh H, et al (2003) Expression of RET in follicular cell-derived tumors of the thyroid gland: prevalence and implication of morphological type. *Pathol Int* 53:146–153
216. Nikiforov YE (2002) RET/PTC rearrangement in thyroid tumors. *Endocr Pathol* 13:3–16
217. Jhiang SM, Cho JY, Furminger TL, et al (1998) Thyroid carcinomas in RET/PTC transgenic mice. *Recent Results Cancer Res* 154:265–270
218. Fischer AH, Bond JA, Taysavang P, Eugene B, Wynford-Thomas D (1998) Papillary thyroid carcinoma oncogene (RET/PTC) alters the nuclear envelope and chromatin structure. *Am J Pathol* 153:1443–1450
219. Fusco A, Santoro M, Grieco M, et al (1995) RET/PTC activation in human thyroid carcinomas. *J Endocrinol Invest* 18:127–129
220. Nikiforov YE, Rowland JM, Bove KE, Monforte-Munoz H, Fagin JA (1997) Distinct pattern of ret oncogene rearrangements in morphological variants of radiation-induced and sporadic thyroid papillary carcinomas in children. *Cancer Res* 57:1690–1694

221. Bounacer A, Wicker R, Schlumberger M, Sarasin A, Suarez HG (1997) Oncogenic rearrangements of the ret proto-oncogene in thyroid tumors induced after exposure to ionizing radiation. *Biochimie* 79:619–623
222. Wirtschafter A, Schmidt R, Rosen D, et al (1997) Expression of the RET/PTC fusion gene as a marker for papillary carcinoma in Hashimoto's thyroiditis [see comments]. *Laryngoscope* 107:95–100
223. Sheils O, Smyth P, Finn S, Sweeney EC, O'Leary JJ (2002) RET/PTC rearrangements in Hashimoto's thyroiditis. *Int J Surg Pathol* 10:167–168; discussion 168–169
224. Elisei R, Romei C, Vorontsova T, et al (2001) RET/PTC rearrangements in thyroid nodules: studies in irradiated and not irradiated, malignant and benign thyroid lesions in children and adults. *J Clin Endocrinol Metab* 86:3211–3216
225. Cohen Y, Xing M, Mambo E, et al (2003) BRAF mutation in papillary thyroid carcinoma. *J Natl Cancer Inst* 95:625–627
226. Nikiforova MN, Kimura ET, Gandhi M, et al (2003) BRAF mutations in thyroid tumors are restricted to papillary carcinomas and anaplastic or poorly differentiated carcinomas arising from papillary carcinomas. *J Clin Endocrinol Metab* 88:5399–5404
227. Soares P, Trovisco V, Rocha AS, et al (2004) BRAF mutations typical of papillary thyroid carcinoma are more frequently detected in undifferentiated than in insular and insular-like poorly differentiated carcinomas. *Virchows Arch* 444:572–576
228. Puxeddu E, Moretti S, Elisei R, et al (2004) BRAF(V599E) mutation is the leading genetic event in adult sporadic papillary thyroid carcinomas. *J Clin Endocrinol Metab* 89:2414–2420
229. Begum S, Rosenbaum E, Henrique R, Cohen Y, Sidransky D, Westra WH (2004) BRAF mutations in anaplastic thyroid carcinoma: implications for tumor origin, diagnosis and treatment. *Mod Pathol* 17:1359–1363
230. Mazzaferri EL (1999) An overview of the management of papillary and follicular thyroid carcinoma. *Thyroid* 9:421–427
231. Kjellman P, Learoyd DL, Messina M, et al (2001) Expression of the RET proto-oncogene in papillary thyroid carcinoma and its correlation with clinical outcome. *Br J Surg* 88:557–563
232. Basolo F, Molinaro E, Agate L, et al (2001) RET protein expression has no prognostic impact on the long-term outcome of papillary thyroid carcinoma. *Eur J Endocrinol* 145:599–604
233. Cetta F, Gori M, Raffaelli N, Baldi C, Montalto G (1999) Comment on clinical and prognostic relevance of Ret-PTC activation in patients with papillary thyroid carcinoma. *J Clin Endocrinol Metab* 84:2257–2258
234. Kim JY, Cho H, Rhee BD, Kim HY (2002) Expression of CD44 and cyclin D1 in fine needle aspiration cytology of papillary thyroid carcinoma. *Acta Cytol* 46:679–683
235. Takeuchi Y, Daa T, Kashima K, Yokoyama S, Nakayama I, Noguchi S (1999) Mutations of p53 in thyroid carcinoma with an insular component. *Thyroid* 9:377–381
236. Basolo F, Caligo MA, Pinchera A, et al (2000) Cyclin D1 overexpression in thyroid carcinomas: relation with clinicopathological parameters, retinoblastoma gene product, and Ki67 labeling index. *Thyroid* 10:741–746
237. Lee A, LiVolsi VA, Baloch ZW (2000) Expression of DNA topoisomerase IIalpha in thyroid neoplasia. *Mod Pathol* 13:396–400
238. Tallini G, Garcia-Rostan G, Herrero A, et al (1999) Down-regulation of p27KIP1 and Ki67/Mib1 labeling index support the classification of thyroid carcinoma into prognostically relevant categories. *Am J Surg Pathol* 23:678–685
239. DeLellis RA, Lloyd RD, Heitz PU, Eng C (eds) (2004) WHO: pathology and genetics. Tumours of endocrine organs. In: Kleihues P, Sobin LE (eds) WHO classification of tumours. IARC Press, Lyon, France
240. Rodriguez JM, Moreno A, Parrilla P, et al (1997) Papillary thyroid microcarcinoma: clinical study and prognosis. *Eur J Surg* 163:255–259
241. Hay ID, Grant CS, van Heerden JA, Goellner JR, Ebersold JR, Bergstralh EJ (1992) Papillary thyroid microcarcinoma: a study of 535 cases observed in a 50-year period. *Surgery* 112:1139–1146; discussion 1146–1147
242. Braga M, Graf H, Ogata A, Batista J, Hakim NC (2002) Aggressive behavior of papillary microcarcinoma in a patient with Graves' disease initially presenting as cystic neck mass. *J Endocrinol Invest* 25:250–253
243. Fernandez-Real JM, Ricart W (1999) Familial papillary thyroid microcarcinoma. *Lancet* 353:1973–1974
244. Lupoli G, Vitale G, Caraglia M, et al (1999) Familial papillary thyroid microcarcinoma: a new clinical entity. *Lancet* 353:637–639
245. Lindsay S (1960) Carcinoma of the thyroid gland: a clinical and pathologic study of 239 patients at the University of California Hospital. Springfield, IL
246. Chen KTC, Rosai J (1977) Follicular variant of thyroid papillary carcinoma: a clinicopathologic study of six cases. *Am J Surg Pathol* 1:123–130
247. LiVolsi VA, Asa SL (1994) The demise of follicular carcinoma of the thyroid gland. *Thyroid* 4:233–236
248. Baloch ZW, Gupta PK, Yu GH, Sack MJ, LiVolsi VA (1999) Follicular variant of papillary carcinoma. Cytologic and histologic correlation. *Am J Clin Pathol* 111:216–222
249. Baloch Z, LiVolsi VA, Henricks WH, Sebak BA (2002) Encapsulated follicular variant of papillary thyroid carcinoma. *Am J Clin Pathol* 118:603–605; discussion 605–606
250. Tielens ET, Sherman SI, Hruban RH, Ladenson PW (1994) Follicular variant of papillary thyroid carcinoma. A clinicopathologic study. *Cancer* 73:424–431
251. Baloch ZW, Livolsi VA (2002) Follicular-patterned lesions of the thyroid: the bane of the pathologist. *Am J Clin Pathol* 117:143–150

252. Mizukami Y, Nonomura A, Michigishi T, Ohmura K, Noguchi M, Ishizaki T (1995) Diffuse follicular variant of papillary carcinoma of the thyroid. *Histopathology* 27:575–577
253. Ivanova R, Soares P, Castro P, Sobrinho-Simoes M (2002) Diffuse (or multinodular) follicular variant of papillary thyroid carcinoma: a clinicopathologic and immunohistochemical analysis of ten cases of an aggressive form of differentiated thyroid carcinoma. *Virchows Arch* 440:418–424
254. Williams ED, Abrosimov A, Bogdanova TI, Roasi J, Sidorov Y, Thomas GA (2000) Two proposals regarding the terminology of thyroid tumors. Guest Editorial. *Int J Surg Pathol* 8:181–183
255. Johnson THLR, Thompson NW, Beierwalters WH, Sisson JC (1988) Prognostic implications of the tall cell variant of papillary carcinoma. *Am J Surg Pathol* 12:22–27
256. Sobrinho-Simoes M, Sambade C, Nesland JM, Johannessen JV (1989) Tall cell papillary carcinoma. *Am J Surg Pathol* 13:79–80
257. Bronner MP, LiVolsi VA (1991) Spindle cell squamous carcinoma of the thyroid: an unusual anaplastic tumor associated with tall cell papillary cancer. *Mod Pathol* 4:637–643
258. Prendiville S, Burman KD, Ringel MD, et al (2000) Tall cell variant: an aggressive form of papillary thyroid carcinoma. *Otolaryngol Head Neck Surg* 122:352–357
259. Jobran R, Baloch ZW, Aviles V, Rosato EF, Schwartz S, LiVolsi VA (2000) Tall cell papillary carcinoma of the thyroid: metastatic to the pancreas [In Process Citation]. *Thyroid* 10:185–187
260. Terry J, St John S, Karkowski F, et al (1994) Tall cell papillary thyroid cancer: incidence and prognosis. *Am J Surg* 168:459–461
261. Sobrinho-Simoes M, Nesland JM, Johannessen JV (1988) Columnar-cell carcinoma. Another variant of poorly differentiated carcinoma of the thyroid. *Am J Clin Pathol* 89:264–267
262. Chan JK (1990) Papillary carcinoma of thyroid: classical and variants. *Histol Histopathol* 5:241–257
263. Wenig BM, Thompson LDR, Adair CF, Shmookler B, Hefess CF (1998) Thyroid papillary carcinoma of columnar cell type. A clinicopathologic study of 16 cases. *Cancer* 82:740–753
264. Evans HL (1996) Encapsulated columnar-cell carcinoma of the thyroid. A report of four cases suggesting a favorable outcome. *Am J Surg Pathol* 20:1205–1211
265. Apel RL, Asa SL, LiVolsi VA (1995) Papillary Hürthle cell carcinoma with lymphocytic stroma. “Warthin-like tumor” of the thyroid. *Am J Surg Pathol* 19:810–814
266. Baloch ZW, LiVolsi VA (2000) Warthin-like papillary carcinoma of the thyroid. *Arch Pathol Lab Med* 124:1192–1195
267. Chan JKC, Tsui MS, Tse CH (1987) Diffuse sclerosing variant of papillary thyroid carcinoma. A histological and immunohistochemical study of three cases. *Histopathology* 11:191–201
268. Peix JL, Mabrut JY, Van Box Som P, Berger N (1998) [Thyroid cancer in children and adolescents. Clinical aspects, diagnostic problems and special therapeutics]. *Ann Endocrinol (Paris)* 59:113–120
269. Santoro M, Thomas GA, Vecchio G, et al (2000) Gene rearrangement and Chernobyl related thyroid cancers. *Br J Cancer* 82:315–322
270. Soares J, Limbert E, Sobrinho-Simoes M (1989) Diffuse sclerosing variant of papillary thyroid carcinoma. A clinicopathologic study of 10 cases. *Pathol Res Pract* 185:200–206
271. Nikiforov YE, Erickson LA, Nikiforova MN, Caudill CM, Lloyd RV (2001) Solid variant of papillary thyroid carcinoma: incidence, clinical-pathologic characteristics, molecular analysis, and biologic behavior. *Am J Surg Pathol* 25:1478–1484
272. Thomas GA, Bunnell H, Cook HA, et al (1999) High prevalence of RET/PTC rearrangements in Ukrainian and Belarussian post-Chernobyl thyroid papillary carcinomas: a strong correlation between RET/PTC3 and the solid-follicular variant. *J Clin Endocrinol Metab* 84:4232–4238
273. Sywak M, Pasiaka JL, Ogilvie T (2004) A review of thyroid cancer with intermediate differentiation. *J Surg Oncol* 86:44–54
274. Vergilio J, Baloch ZW, LiVolsi VA (2002) Spindle cell metaplasia of the thyroid arising in association with papillary carcinoma and follicular adenoma. *Am J Clin Pathol* 117:199–204
275. Dickersin G, Vickery AL Jr, Smith S (1980) Papillary carcinoma of the thyroid, oxyphil cell type, “clear cell” variant: a light and electron microscopic study. *Am J Surg Pathol* 4:501–509
276. Berho M, Suster S (1997) The oncocytic variant of papillary carcinoma of the thyroid: a clinicopathologic study of 15 cases. *Hum Pathol* 28:47–53
277. Bisi H, Longatto Filho A, de Camargo RY, Fernandes VS (1993) Thyroid papillary carcinoma lipomatous type: report of two cases. *Pathologica* 85:761–764
278. Schroder S, Bocker W (1985) Lipomatous lesions of the thyroid gland: a review. *Appl Pathol* 3:140–149
279. Chan JK, Carcangiu ML, Rosai J (1991) Papillary carcinoma of thyroid with exuberant nodular fasciitis-like stroma. Report of three cases. *Am J Clin Pathol* 95:309–314
280. Cameselle-Teijeiro J, Chan JK (1999) Cribriform-morular variant of papillary carcinoma: a distinctive variant representing the sporadic counterpart of familial adenomatous polyposis-associated thyroid carcinoma? *Mod Pathol* 12:400–411

281. Hirokawa M, Kuma S, Miyauchi A, et al (2004) Morules in cribriform-morular variant of papillary thyroid carcinoma: immunohistochemical characteristics and distinction from squamous metaplasia. *APMIS* 112:275–282
282. Xu B, Yoshimoto K, Miyauchi A, et al (2003) Cribriform-morular variant of papillary thyroid carcinoma: a pathological and molecular genetic study with evidence of frequent somatic mutations in exon 3 of the beta-catenin gene. *J Pathol* 199:58–67
283. Franssila KO, Ackerman LV, Brown CL, Hedinger CE (1985) Follicular carcinoma. *Semin Diagn Pathol* 2:101–122
284. Tollefson HR, Shah JP, Huvos AG (1973) Follicular carcinoma of the thyroid. *Am J Surg* 126:523–528
285. Wade JS (1975) The aetiology and diagnosis of malignant tumours of the thyroid gland. *Br J Surg* 62:760–764
286. Jorda M, Gonzalez-Campora R, Mora J, Herrero-Zapatero A, Otal C, Galera H (1993) Prognostic factors in follicular carcinoma of the thyroid. *Arch Pathol Lab Med* 117:631–635
287. Segal K, Arad A, Lubin E, Shpitzer T, Hadar T, Feinmesser R (1994) Follicular carcinoma of the thyroid. *Head Neck* 16:533–538
288. Crile G, Pontius K, Hawk W (1985) Factors influencing the survival of patients with follicular carcinoma of the thyroid gland. *Surg Gynecol Obstet* 160:409–412
289. Schmidt RJ, Wang CA (1986) Encapsulated follicular carcinoma of the thyroid: diagnosis, treatment, and results. *Surgery* 100:1068–1077
290. Evans HL (1984) Follicular neoplasms of the thyroid. A study of 44 cases followed for a minimum of 10 years with emphasis on differential diagnosis. *Cancer* 54:535–540
291. Thompson LD, Wieneke JA, Paal E, Frommelt RA, Adair CF, Heffess CS (2001) A clinicopathologic study of minimally invasive follicular carcinoma of the thyroid gland with a review of the English literature. *Cancer* 91:505–524
292. Carcangiu ML (1997) Minimally invasive follicular carcinoma. *Endocr Pathol* 8:231–234
293. Shaha AR, Loree TR, Shah JP (1995) Prognostic factors and risk group analysis in follicular carcinoma of the thyroid. *Surgery* 118:1131–1136; discussion 1136–1138
294. Moore JH Jr, Bacharach B, Choi HY (1985) Anaplastic transformation of metastatic follicular carcinoma of the thyroid. *J Surg Oncol* 29:216–221
295. D'Avanzo A, Treseler P, Ituarte PH, et al (2004) Follicular thyroid carcinoma: histology and prognosis. *Cancer* 100:1123–1129
296. Jakubiak-Wielganowicz M, Kubiak R, Sygut J, Pomorski L, Kordek R (2003) Usefulness of galectin-3 immunohistochemistry in differential diagnosis between thyroid follicular carcinoma and follicular adenoma. *Pol J Pathol* 54:111–115
297. Collini P, Sampietro G, Rosai J, Pilotti S (2003) Minimally invasive (encapsulated) follicular carcinoma of the thyroid gland is the low-risk counterpart of widely invasive follicular carcinoma but not of insular carcinoma. *Virchows Arch* 442:71–76
298. Kahn NF, Perzin KH (1983) Follicular carcinoma of the thyroid: an evaluation of the histologic criteria used for diagnosis. *Pathol Ann* 18:221–253
299. Udelsman R, Westra WH, Donovan PI, Sohn TA, Cameron JL (2001) Randomized prospective evaluation of frozen-section analysis for follicular neoplasms of the thyroid. *Ann Surg* 233:716–722
300. Leteurtre E, Leroy X, Pattou F, et al (2001) Why do frozen sections have limited value in encapsulated or minimally invasive follicular carcinoma of the thyroid? *Am J Clin Pathol* 115:370–374
301. Paphavasit A, Thompson GB, Hay ID, et al (1997) Follicular and Hürthle cell thyroid neoplasms. Is frozen-section evaluation worthwhile? *Arch Surg* 132:674–678; discussion 678–680
302. Johannessen JV, Sobrinho-Simoes M (1982) Follicular carcinoma of the human thyroid gland. An ultrastructural study with emphasis on scanning electron microscopy. *Diagn Histopathol* 5:113–127
303. Backdahl M (1985) Nuclear DNA content and prognosis in papillary, follicular, and medullary carcinomas of the thyroid. Doctoral thesis, Karolinska Medical Institute, Stockholm, Sweden
304. Papotti M, Rodriguez J, Pompa RD, Bartolazzi A, Rosai J (2004) Galectin-3 and HBME-1 expression in well-differentiated thyroid tumors with follicular architecture of uncertain malignant potential. *Mod Pathol* 18:541–546
305. Rosai J (2003) Immunohistochemical markers of thyroid tumors: significance and diagnostic applications. *Tumori* 89:517–519
306. Kroll TG, Sarraf P, Pecciarini L, et al (2000) PAX8–PPAR[gamma] 1 fusion in oncogene human thyroid carcinoma. *Science* 289:1357–1360
307. Marques AR, Espadinha C, Catarino AL, et al (2002) Expression of PAX8–PPAR gamma 1 rearrangements in both follicular thyroid carcinomas and adenomas. *J Clin Endocrinol Metab* 87:3947–3952
308. Gustafson KS, LiVolsi VA, Furth EE, Pasha TL, Putt ME, Baloch ZW (2003) Peroxisome proliferator-activated receptor gamma expression in follicular-patterned thyroid lesions. Caveats for the use of immunohistochemical studies. *Am J Clin Pathol* 120:175–181
309. Esapa CT, Johnson SJ, Kendall-Taylor P, Lennard TW, Harris PE (1999) Prevalence of Ras mutations in thyroid neoplasia. *Clin Endocrinol* 50:529–535
310. Basolo F, Pisaturo F, Pollina LE, et al (2000) N-ras mutation in poorly differentiated thyroid carcinomas: correlation with bone metastases and inverse correlation to thyroglobulin expression. *Thyroid* 10:19–23



311. Capella G, Matias-Guiu X, Ampudia X, de Leiva A, Perucho M, Prat J (1996) Ras oncogene mutations in thyroid tumors: polymerase chain reaction-restriction-fragment-length polymorphism analysis from paraffin-embedded tissues. *Diagn Mol Pathol* 5:45–52
312. Grebe SK, McIver B, Hay ID, et al (1997) Frequent loss of heterozygosity on chromosomes 3p and 17p without VHL or p53 mutations suggests involvement of unidentified tumor suppressor genes in follicular thyroid carcinoma. *J Clin Endocrinol Metab* 82:3684–3691
313. Matsuo K, Tang SH, Fagin JA (1991) Allelotype of human thyroid tumors: loss of chromosome 11q13 sequences in follicular neoplasms. *Mol Endocrinol* 5:1873–1879
314. Nesland JM, Sobrinho-Simoes MA, Holm R, Sambade MC, Johannessen JV (1985) Hürthle-cell lesions of the thyroid: a combined study using transmission electron microscopy, scanning electron microscopy, and immunocytochemistry. *Ultrastruct Pathol* 8:269–290
315. Gonzalez-Campora R, Herrero-Zapatero A, Lerma E, Sanchez F, Galera H (1986) Hürthle cell and mitochondrion-rich cell tumors. A clinicopathologic study. *Cancer* 57:1154–1163
316. Thompson N, Dun E, Batsakis J, Nishiyama R (1974) Hürthle cell lesions of the thyroid gland. *Surg Gynecol Obstet* 139:555–560
317. Gundry S, Burney R, Thompson N, Lloyd R (1983) Total thyroidectomy for Hürthle cell neoplasm of the thyroid gland. *Arch Surg* 118:529–553
318. Bronner M, LiVolsi V (1988) Oxyphilic (Askanazy/Hürthle cell) tumors of the thyroid: microscopic features predict biologic behavior. *Surg Pathol* 1:137–150
319. Carcangiu ML, Bianchi S, Savino D, Voynick IM, Rosai J (1991) Follicular Hürthle cell tumors of the thyroid gland. *Cancer* 68:1944–1953
320. Chen H, Nicol TL, Zeiger MA, et al (1998) Hürthle cell neoplasms of the thyroid: are there factors predictive of malignancy? *Ann Surg* 227:542–546
321. Janser JC, Pusel J, Rodier JE, Navarrete E, Rodier D (1989) [Hürthle cell tumor of the thyroid gland. Analysis of a series of 33 cases]. *J Chir (Paris)* 126:619–624
322. Kanthan R, Radhi JM (1998) Immunohistochemical analysis of thyroid adenomas with Hürthle cells. *Pathology* 30:4–6
323. Bronner MP, Clevenger CV, Edmonds PR, Lowell DM, McFarland MM, LiVolsi VA (1988) Flow cytometric analysis of DNA content in Hürthle cell adenomas and carcinomas of the thyroid. *Am J Clin Pathol* 89:764–769
324. Scharck C, Fulton N, Yashiro T, et al (1992) The value of measurement of ras oncogenes and nuclear DNA analysis in the diagnosis of Hürthle cell tumors of the thyroid. *World J Surg* 16:745–751; discussion 752
325. Bouras M, Bertholon J, Dutrieux-Berger N, Parvaz P, Paulin C, Revol A (1998) Variability of Ha-ras (codon 12) proto-oncogene mutations in diverse thyroid cancers. *Eur J Endocrinol* 139:209–216
326. Maximo V, Soares P, Lima J, Cameselle-Teijeiro J, Sobrinho-Simoes M (2002) Mitochondrial DNA somatic mutations (point mutations and large deletions) and mitochondrial DNA variants in human thyroid pathology: a study with emphasis on Hürthle cell tumors. *Am J Pathol* 160:1857–1865
327. Variakojis D, Getz ML, Paloyan E, Straus FH (1975) Papillary clear cell carcinoma of the thyroid gland. *Hum Pathol* 6:384–390
328. Carcangiu ML, Sibley RK, Rosai J (1985) Clear cell change in primary thyroid tumors. A study of 38 cases. *Am J Surg Pathol* 9:705–722
329. Lam KY, Lo CY (1998) Metastatic tumors of the thyroid gland: a study of 79 cases in Chinese patients. *Arch Pathol Lab Med* 122:37–41
330. Sakamoto A, Kasai N, Sugano H (1983) Poorly differentiated carcinoma of the thyroid. *Cancer* 52:1849–1855
331. Carcangiu ML, Zampi G, Rosai J (1984) Poorly differentiated (“insular”) thyroid carcinoma. A reinterpretation of Langhans’ “wuchernde Struma.” *Am J Surg Pathol* 8:655–668
332. Flynn SD, Forman BH, Stewart AF, Kinder BK (1988) Poorly differentiated (“insular”) carcinoma of the thyroid gland: an aggressive subset of differentiated thyroid neoplasms. *Surgery* 104:963–970
333. Palestini N, Papotti M, Durando R, Fortunato MA (1993) [Poorly differentiated “insular” carcinoma of the thyroid: long-term survival]. *Minerva Chir* 48:1301–1305
334. Nishida T, Katayama S, Tsujimoto M, Nakamura J, Matsuda H (1999) Clinicopathological significance of poorly differentiated thyroid carcinoma. *Am J Surg Pathol* 23:205–211
335. Venkatesh YS, Ordonez NG, Schultz PN, Hickey RC, Gopfert H, Samaan NA (1990) Anaplastic carcinoma of the thyroid. A clinicopathologic study of 121 cases. *Cancer* 66:321–330
336. Dumitriu L, Stefanescu L, Tasca C (1984) The anaplastic transformation of differentiated thyroid carcinoma. An ultrastructural study. *Endocrinologie* 22:91–96
337. Chang TC, Liaw KY, Kuo SH, Chang CC, Chen FW (1989) Anaplastic thyroid carcinoma: review of 24 cases, with emphasis on cytodagnosis and leukocytosis. *Taiwan Yi Xue Hui Za Zhi* 88:551–556
338. LiVolsi VA, Brooks JJ, Arendash-Durand B (1987) Anaplastic thyroid tumors. *Immunohistology*. *Am J Clin Pathol* 87:434–442
339. Berry B, MacFarlane J, Chan N (1990) Osteoclastoma-like anaplastic carcinoma of the thyroid. Diagnosis by fine needle aspiration cytology. *Acta Cytol* 34:248–250
340. Wan SK, Chan JK, Tang SK (1996) Paucicellular variant of anaplastic thyroid carcinoma. A mimic of Riedel’s thyroiditis. *Am J Clin Pathol* 105:388–393
341. Giuffrida D, Attard M, Marasa L, et al (2000) Thyroid carcinosarcoma, a rare and aggressive histotype: a case report. *Ann Oncol* 11:1497–1499

342. Donnell CA, Pollock WJ, Sybers WA (1987) Thyroid carcinosarcoma. *Arch Pathol Lab Med* 111:1169–1172
343. Miettinen M, Franssila KO (2000) Variable expression of keratins and nearly uniform lack of thyroid transcription factor 1 in thyroid anaplastic carcinoma. *Hum Pathol* 31:1139–1145
344. Neri A, Aldovini D, Leonardi E, Giampiccolo M, Pedrolli C (1990) [Primary angiosarcoma of the thyroid gland. Presentation of a clinical case]. *Recenti Prog Med* 81:318–321
345. Tsugawa K, Koyanagi N, Nakanishi K, et al (1999) Leiomyosarcoma of the thyroid gland with rapid growth and tracheal obstruction: a partial thyroidectomy and tracheostomy using an ultrasonically activated scalpel can be safely performed with less bleeding. *Eur J Med Res* 4:483–487
346. Chan YF, Ma L, Boey JH, Yeung HY (1986) Angiosarcoma of the thyroid. An immunohistochemical and ultrastructural study of a case in a Chinese patient. *Cancer* 57:2381–2388
347. Sahoo M, Bal CS, Bhatnagar D (2002) Primary squamous-cell carcinoma of the thyroid gland: new evidence in support of follicular epithelial cell origin. *Diagn Cytopathol* 27:227–231
348. Harach HR, Day ES, de Strizic NA (1986) Mucoepidermoid carcinoma of the thyroid. Report of a case with immunohistochemical studies. *Medicina* 46:213–216
349. Arezzo A, Patetta R, Ceppa P, Borgonovo G, Torre G, Mattioli FP (1998) Mucoepidermoid carcinoma of the thyroid gland arising from a papillary epithelial neoplasm. *Am Surg* 64:307–311
350. Wenig BM, Adair CF, Heffess CS (1995) Primary mucoepidermoid carcinoma of the thyroid gland: a report of six cases and a review of the literature of a follicular epithelial-derived tumor. *Hum Pathol* 26:1099–1108
351. Chan JK, Albores-Saavedra J, Battifora H, Carcangiu ML, Rosai J (1991) Sclerosing mucoepidermoid thyroid carcinoma with eosinophilia. A distinctive low-grade malignancy arising from the metaplastic follicles of Hashimoto's thyroiditis. *Am J Surg Pathol* 15:438–448
352. Chan JK, Rosai J (1991) Tumors of the neck showing thymic or related branchial pouch differentiation: a unifying concept. *Hum Pathol* 22:349–367
353. Iwasa K, Imai MA, Noguchi M, et al (2002) Spindle epithelial tumor with thymus-like differentiation (SETTLE) of the thyroid. *Head Neck* 24:888–893
354. Ahuja AT, Chan ES, Allen PW, Lau KY, King W, Metreweli C (1998) Carcinoma showing thymiclike differentiation (CASTLE tumor). *AJNR Am J Neuroradiol* 19:1225–1228
355. Bayer-Garner IB, Kozovska ME, Schwartz MR, Reed JA (2004) Carcinoma with thymus-like differentiation arising in the dermis of the head and neck. *J Cutan Pathol* 31:625–629
356. Roka S, Kornek G, Schuller J, Ortman E, Feichtinger J, Armbruster C (2004) Carcinoma showing thymic-like elements: a rare malignancy of the thyroid gland. *Br J Surg* 91:142–145
357. Hazard JB, Hawk WA, Crile G (1959) Medullary (solid) carcinoma of the thyroid. A clinicopathologic entity. *J Clin Endocrinol Metab* 19:152–161
358. Williams ED (1965) A review of 17 cases of carcinoma of the thyroid and pheochromocytoma. *J Clin Pathol* 18:288–292
359. Williams ED (1966) Histogenesis of medullary carcinoma of the thyroid. *J Clin Pathol* 19:114–118
360. Block MA, Horn RC, Miller JM, Barrett JL, Brush BE (1967) Familial medullary carcinoma of the thyroid. *Ann Surg* 166:403–412
361. Albores-Saavedra J, LiVolsi VA, Williams ED (1985) Medullary carcinoma. *Semin Diagn Pathol* 2:137–146
362. Wolfe HJ, Melvin KE, Cervi-Skinner SJ, et al (1973) C-cell hyperplasia preceding medullary thyroid carcinoma. *N Engl J Med* 289:437–441
363. Mulligan LM, Kwok JB, Healey CS, et al (1993) Germ-line mutations of the RET proto-oncogene in multiple endocrine neoplasia type 2A. *Nature* 363:458–460
364. Hofstra RM, Landsvater RM, Ceccherini I, et al (1994) A mutation in the RET proto-oncogene associated with multiple endocrine neoplasia type 2B and sporadic medullary thyroid carcinoma [see comments]. *Nature* 367:375–376
365. Mulligan LM, Eng C, Healey CS, et al (1994) Specific mutations of the RET proto-oncogene are related to disease phenotype in MEN 2A and FMTC. *Nat Genet* 6:70–74
366. Uribe M, Fenoglio-Preiser CM, Grimes M, Feind C (1985) Medullary carcinoma of the thyroid gland. Clinical, pathological and immunohistochemical features with review of the literature. *Am J Surg Pathol* 9:577–594
367. Wolfe HJ, Delellis RA (1981) Familial medullary thyroid carcinoma and C cell hyperplasia. *Clin Endocrinol Metab* 10:351–365
368. Leboulleux S, Baudin E, Travagli JP, Schlumberger M (2004) Medullary thyroid carcinoma. *Clin Endocrinol (Oxf)* 61:299–310
369. Williams E, Karim S, Sandler M (1968) Prostaglandin secretion by medullary carcinoma of the thyroid: a possible cause of the associated diarrhea. *Lancet* 1:22–23
370. Kakudo K, Miyauchi A, Ogihara T, et al (1982) Medullary carcinoma of the thyroid with ectopic ACTH syndrome. *Acta Pathol Jpn* 32:793–800
371. Sipple JH (1961) The association of pheochromocytoma with carcinoma of the thyroid gland. *Am J Med* 31:163–166
372. Jansson S, Hansson G, Salander H, Stenstrom G, Tisell L (1984) Prevalence of C-cell hyperplasia and medullary thyroid carcinoma in a consecutive series of pheochromocytoma patients. *World J Surg* 8:493–500

373. Eng C (1996) RET proto-oncogene in multiple endocrine neoplasia type 2 and Hirschprung's disease. *Semin Med Beth Israel Hosp, Boston* 335:943–951
374. Eng C (1999) RET proto-oncogene in the development of human cancer. *J Clin Oncol* 17:380–393
375. Eng C, Clayton D, Schuffenecker I, et al (1996) The relationship between specific RET proto-oncogene mutations and disease phenotype in multiple endocrine neoplasia type 2. International RET mutation consortium analysis. *JAMA* 276:1575–1579
376. Kebebew E, Ituarte PH, Siperstein AE, Duh QY, Clark OH (2000) Medullary thyroid carcinoma: clinical characteristics, treatment, prognostic factors, and a comparison of staging systems. *Cancer* 88:1139–1148
377. Kambouris M, Jackson CE, Feldman GL (1996) Diagnosis of multiple endocrine neoplasia (MEN) 2A, 2B and familial medullary thyroid cancer (FMTC) by multiplex PCR and heteroduplex analyses of RET proto-oncogene mutations. *Hum Mutat* 8:64–70
378. Nakata S, Okugi H, Saitoh Y, Takahashi H, Shimizu K (2001) Multiple endocrine neoplasia type 2B. *Int J Urol* 8:398–400
379. Nguyen L, Niccoli-Sire P, Caron P, et al (2001) Pheochromocytoma in multiple endocrine neoplasia type 2: a prospective study. *Eur J Endocrinol* 144:37–44
380. Marsh DJ, Zheng Z, Arnold A, et al (1997) Mutation analysis of glial cell line-derived neurotrophic factor, a ligand for an RET/coreceptor complex, in multiple endocrine neoplasia type 2 and sporadic neuroendocrine tumors. *J Clin Endocrinol Metab* 82:3025–3028
381. Borrello MG, Smith DP, Pasini B, et al (1995) RET activation by germline MEN2A and MEN2B mutations. *Oncogene* 11:2419–2427
382. Eng C, Smith DP, Mulligan LM, et al (1994) Point mutation within the tyrosine kinase domain of the RET proto-oncogene in multiple endocrine neoplasia type 2B and related sporadic tumours. *Hum Mol Genet* 3:237–241
383. Cohen EG, Shaha AR, Rinaldo A, Devaney KO, Ferlito A (2004) Medullary thyroid carcinoma. *Acta Otolaryngol* 124:544–557
384. Asa SL (1997) C-cell lesions of the thyroid. *Pathol Case Rev* 2:210–217
385. Abrosimov A (1996) [Histologic and immunohistochemical characterization of medullary thyroid carcinoma]. *Arkh Patol* 58:43–48
386. Alevizaki M, Dai K, Grigorakis SI, Legon S, Souvatzoglou A (1994) Amylin/islet amyloid polypeptide expression in medullary carcinoma of the thyroid: correlation with the expression of the related calcitonin/CGRP genes. *Clin Endocrinol (Oxf)* 41:21–26
387. Dominguez-Malagon H, Delgado-Chavez R, Torres-Najera M, Gould E, Albores-Saavedra J (1989) Oxyphil and squamous variants of medullary thyroid carcinoma. *Cancer* 63:1183–1188
388. Harach HR, Williams ED (1983) Glandular (tubular and follicular) variants of medullary carcinoma of the thyroid. *Histopathology* 7:83–97
389. Huss LJ, Mendelsohn G (1990) Medullary carcinoma of the thyroid gland: an encapsulated variant resembling the hyalinizing trabecular (paraganglioma-like) adenoma of thyroid. *Mod Pathol* 3:581–585
390. Landon G, Ordonez NG (1985) Clear cell variant of medullary carcinoma of the thyroid. *Hum Pathol* 16:844
391. Mendelsohn G, Baylin SB, Bigner SH, Wells SA, Jr., Eggleston JC (1980) Anaplastic variants of medullary thyroid carcinoma: a light-microscopic and immunohistochemical study. *Am J Surg Pathol* 4:333–341
392. Kos M, Separovic V, Sarcevic B (1995) Medullary carcinoma of the thyroid: histomorphological, histochemical and immunohistochemical analysis of twenty cases. *Acta Med Croatica* 49:195–199
393. DeLilles RA, Rule AH, Spiler F, et al (1978) Calcitonin and carcinoembryonic antigen as tumor markers in medullary thyroid carcinoma. *Am J Clin Pathol* 70:587
394. Hirsch MS, Faquin WC, Krane JF (2004) Thyroid transcription factor-1, but not p53, is helpful in distinguishing moderately differentiated neuroendocrine carcinoma of the larynx from medullary carcinoma of the thyroid. *Mod Pathol* 17:631–636
395. Matsubayashi S, Yanaihara C, Ohkubo M, et al (1984) Gastrin-releasing peptide immunoreactivity in medullary thyroid carcinoma. *Cancer* 53:2472
396. Roth KA, Bensch KG, Hoffman AR (1987) Characterization of opioid peptides in human thyroid medullary carcinoma. *Cancer* 59:1594
397. Komminoth P, Roth J, Saremasiani P, et al (1994) Polysialic acid of the neural cell adhesion molecule in the human thyroid: a marker for medullary carcinoma and primary C-cell hyperplasia. An immunohistochemical study on 79 thyroid lesions. *Am J Surg Pathol* 18:399
398. Ruppert JM, Eggleston JC, deBustros A, Baylin SB (1986) Disseminated calcitonin-poor medullary thyroid carcinoma in a patient with calcitonin-rich primary tumor. *Am J Surg Pathol* 10:513–518
399. Randolph GW, Maniar D (2000) Medullary carcinoma of the thyroid. *Cancer Control* 7:253–261
400. Giuffrida D, Ferrau F, Bordonaro R, et al (2000) [Medullary carcinoma of the thyroid: diagnosis and therapy]. *Clin Ter* 151:29–35
401. Gimm O, Sutter T, Dralle H (2001) Diagnosis and therapy of sporadic and familial medullary thyroid carcinoma. *J Cancer Res Clin Oncol* 127:156–165
402. Gilliland FD, Hunt WC, Morris DM, Key CR (1997) Prognostic factors for thyroid carcinoma. A population-based study of 15,698 cases from the Surveillance, Epidemiology and End Results (SEER) program 1973–1991. *Cancer* 79:564–573

403. Randolph GW (1996) Medullary carcinoma of the thyroid: subtypes and current management. *Compr Ther* 22:203–210
404. Brierley J, Tsang R, Simpson WJ, Gospodarowicz M, Sutcliffe S, Panzarella T (1996) Medullary thyroid cancer: analyses of survival and prognostic factors and the role of radiation therapy in local control. *Thyroid* 6:305–310
405. Schroder S, Bocker W, Baisch H, et al (1988) Prognostic factors in medullary thyroid carcinomas. Survival in relation to age, sex, stage, histology, immunocytochemistry, and DNA content. *Cancer* 61:806–816
406. Albores-Saavedra J, Gorraez de la Mora T, de la Torre-Rendon F, Gould E (1990) Mixed medullary-papillary carcinoma of the thyroid: a previously unrecognized variant of thyroid carcinoma. *Hum Pathol* 21:1151–1155
407. Giove E, Renzulli G, Lorusso C, Merlicco D, Iacobone D (2004) [Mixed medullary and follicular carcinoma of the thyroid: report of one case]. *Ann Ital Chir* 75:251–256; discussion 257
408. Kashima K, Yokoyama S, Inoue S, et al (1993) Mixed medullary and follicular carcinoma of the thyroid: report of two cases with an immunohistochemical study. *Acta Pathol Jpn* 43:428–433
409. LiVolsi VA (2004) Mixed follicular medullary thyroid carcinoma. *Diagn Cytopathol* 31:434; author reply 435
410. Beressi N, Campos JM, Beressi JP, et al (1998) Sporadic medullary microcarcinoma of the thyroid: a retrospective analysis of eighty cases. *Thyroid* 8:1039–1044
411. Guyétant S, Dupre F, Bigorgne JC, et al (1999) Medullary thyroid microcarcinoma: a clinicopathologic retrospective study of 38 patients with no prior familial disease. *Hum Pathol* 30:957–963
412. Albores-Saavedra JA, Krueger JE (2001) C-cell hyperplasia and medullary thyroid microcarcinoma. *Endocr Pathol* 12:365–377
413. Kaserer K, Scheuba C, Neuhold N, et al (2001) Sporadic versus familial medullary thyroid microcarcinoma: a histopathologic study of 50 consecutive patients. *Am J Surg Pathol* 25:1245–1251
414. Mizukami Y, Kurumaya H, Nonomura A, et al (1992) Sporadic medullary microcarcinoma of the thyroid. *Histopathology* 21:375–377
415. Sironi M, Cozzi L, Pareschi R, Spreafico GL, Assi A (1999) Occult sporadic medullary microcarcinoma with lymph node metastases. *Diagn Cytopathol* 21:203–206
416. Russo F, Barone Adesi TL, Arturi A, et al (1997) [Clinicopathological study of microcarcinoma of the thyroid]. *Minerva Chir* 52:891–900
417. Yamauchi A, Tomita Y, Takakuwa T, et al (2002) Polymerase chain reaction-based clonality analysis in thyroid lymphoma. *Int J Mol Med* 10:113–117
418. Ghazanfar S, Quraishy MS, Essa K, Muzaffar S, Saeed MU, Sultan T (2002) Mucosa associated lymphoid tissue lymphoma (MALToma) in patients with cold nodule thyroid. *J Pak Med Assoc* 52:131–133
419. Takano T, Miyauchi A, Matsuzuka F, Yoshida H, Kuma K, Amino N (2000) Diagnosis of thyroid malignant lymphoma by reverse transcription-polymerase chain reaction detecting the monoclonality of immunoglobulin heavy chain messenger ribonucleic acid. *J Clin Endocrinol Metab* 85:671–675
420. Diaz-Arias AA, Bickel JT, Loy TS, Croll GH, Puckett CL, Havey AD (1992) Follicular carcinoma with clear cell change arising in lingual thyroid. *Oral Surg Oral Med Oral Pathol* 74:206–211
421. LiVolsi VA, Perzin KH, Savetsky L (1974) Carcinoma arising in median ectopic thyroid (including thyroglossal duct tissue). *Cancer* 34:1303–1315
422. Doshi SV, Cruz RM, Hilsinger RL Jr (2001) Thyroglossal duct carcinoma: a large case series. *Ann Otol Rhinol Laryngol* 110:734–738
423. Cignarelli M, Ambrosi A, Marino A, Lamacchia O, Cincione R, Neri V (2002) Three cases of papillary carcinoma and three of adenoma in thyroglossal duct cysts: clinical-diagnostic comparison with benign thyroglossal duct cysts. *J Endocrinol Invest* 25:947–954
424. Fih J, Moore R (1963) Ectopic thyroid tissue and ectopic thyroid carcinoma. *Ann Surg* 157:212–222
425. Devaney K, Snyder R, Norris HJ, Tavassoli FA (1993) Proliferative and histologically malignant struma ovarii: a clinicopathologic study of 54 cases. *Int J Gynecol Pathol* 12:333–343
426. Kdous M, Hachicha R, Gamoudi A, et al (2003) [Struma ovarii. Analysis of a series of 7 cases and review of the literature]. *Tunis Med* 81:571–576
427. Rosenblum NG, LiVolsi VA, Edmonds PR, Mikuta JJ (1989) Malignant struma ovarii. *Gynecol Oncol* 32:224–227
428. Koo HL, Jang J, Hong SJ, Shong Y, Gong G (2004) Renal cell carcinoma metastatic to follicular adenoma of the thyroid gland. A case report. *Acta Cytol* 48:64–68
429. Matias-Guiu X, LaGuetta J, Puras-Gil AM, Rosai J (1997) Metastatic neuroendocrine tumors to the thyroid gland mimicking medullary carcinoma: a pathologic and immunohistochemical study of six cases. *Am J Surg Pathol* 21:754–762
430. Baloch ZW, LiVolsi VA (1999) Tumor-to-tumor metastasis to follicular variant of papillary carcinoma of thyroid. *Arch Pathol Lab Med* 123:703–706
431. Bronner MP HR, LiVolsi VA (1994) Utility of frozen section analysis on follicular lesions of the thyroid. *Endocr Pathol* 5:154–161
432. Paessler M, LiVolsi VA, Baloch Z (2001) Role of Ultrafast Papanicolaou stained scrape preparations as an adjunct to frozen section in the surgical management of thyroid lesions. *Endocr Pract* 7:89–94
433. Rodriguez JM, Parrilla P, Sola J, et al (1994) Comparison between preoperative cytology and intraoperative frozen-section biopsy in the diagnosis of thyroid nodules. *Br J Surg* 81:1151–1154

434. Taneri F, Poyraz A, Tekin E, Ersoy E, Dursun A (1998) Accuracy and significance of fine-needle aspiration cytology and frozen section in thyroid surgery. *Endocr Regul* 32:187–191
435. Shaha AR, DiMaio T, Webber C, Jaffe BM (1990) Intraoperative decision making during thyroid surgery based on the results of preoperative needle biopsy and frozen section. *Surgery* 108:964–967; discussion 970–971
436. Shaha A, Gleich L, Di Maio T, Jaffe BM (1990) Accuracy and pitfalls of frozen section during thyroid surgery. *J Surg Oncol* 44:84–92
437. Basolo F, Baloch ZW, Baldanzi A, Miccoli P, LiVolsi VA (1999) Usefulness of Ultrafast Papanicolaou-stained scrape preparations in intraoperative management of thyroid lesions. *Mod Pathol* 12:653–657